Lymph Node FNA

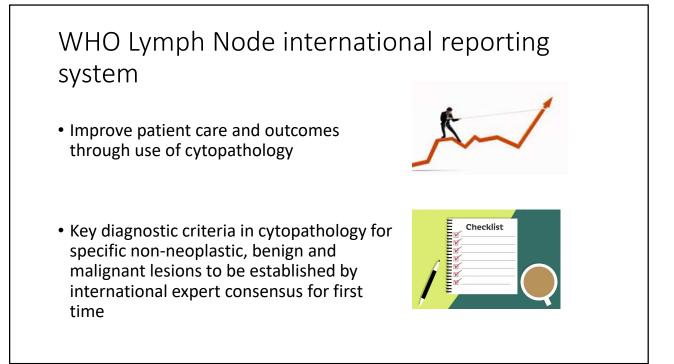
Amy Ly MD Massachusetts General Hospital

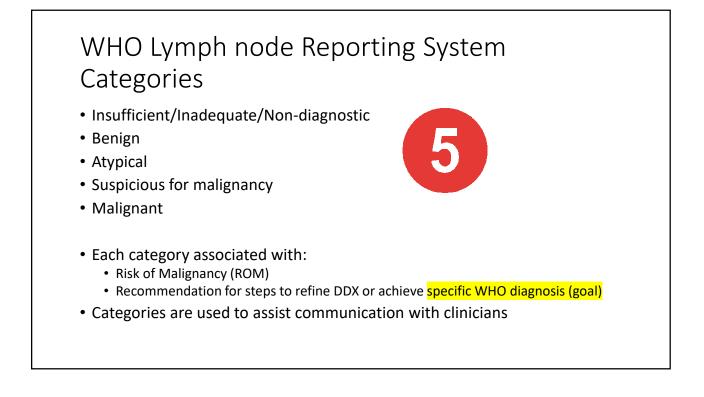
Outline

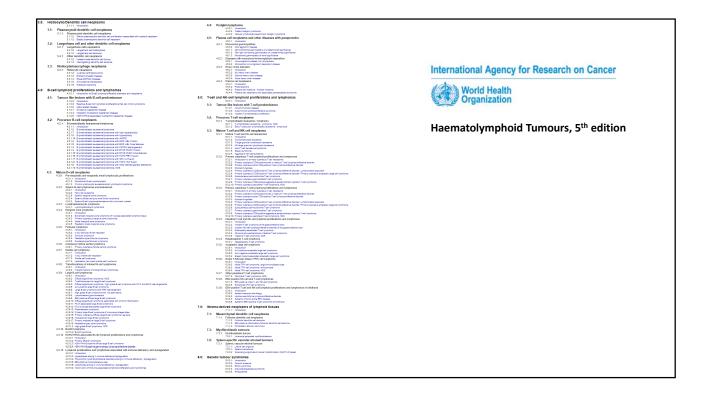
- Preview of structured reporting system for lymph node cytology
- Diagnostic pattern-based approach with brief case presentations

WHO Structured Reporting System (A Preview)

	Acta Cytologica	Fine Needle Aspiration	Fine Needle Aspiration			
	,	Acta Cytologica 2020;64:306-322 DOI: 10.1159/000506497	Received: February 3, 2020 Accepted: February 6, 2020 Published online: May 26, 2020			
	Reporting of Lyn	ne Performance, Classifica nph Node Fine-Needle As _l The Sydney System				
	Maria Calaminici ^d Nancy Mats Ehinger ^h Andrew S.	ena Barroca ^b Beata Bode-Lesniewska ^c P. Caraway ^e David F. Chhieng ^f Immacc Field ^{i-k} William R. Geddie ^{I, m} Ruth L. Ka E. Monaco ^q Arvind Rajwanshi ^r Fernand ^u	atz ⁿ Oscar Lin ^o			
Internationa	Agency for Research o					
Worl	d Health		g System for Lymph Node, Spleen Thymus Cytopathology			

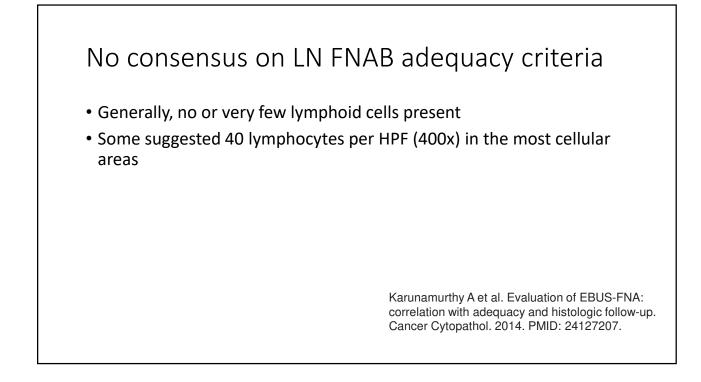


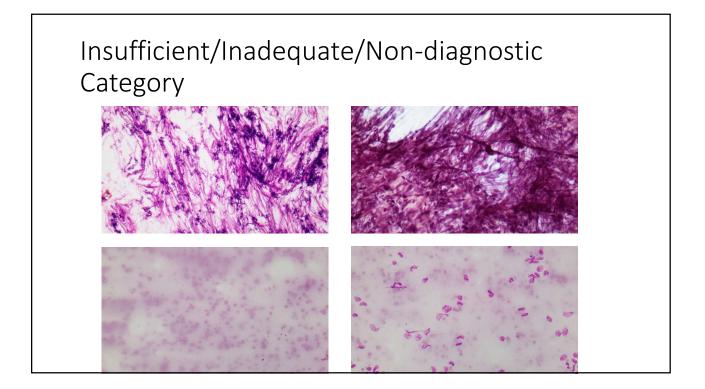




Insufficient/Inadequate/Non-diagnostic Category

- Reliable interpretation cannot be made (qualitative or quantitative limitations)
- E.g. insufficient cellularity, poor smearing technique, air-dry artifact, fixation artifact, obscuring material
- Repeat FNAB is recommended, with ROSE if possible, and with core needle biopsy if available
- If insufficient at time of ROSE, needle rinsings may enable diagnosis when analyzed by flow cytometry, cell block with ICC, cytogenetics, FISH etc



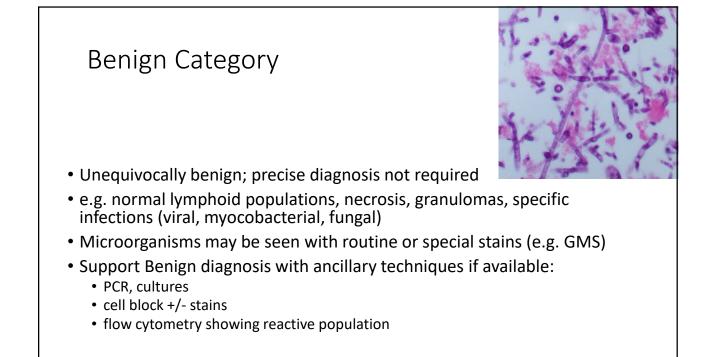


Insufficient/Inadequate/Non-diagnostic Category

- Use one term consistently for clear communication.
- Triple Test: always correlate with imaging and clinical findings.
- WHO system accepts ND diagnosis in cases where there is good lymphoid material, but clinical findings are not explained
 - Others may call these Benign, with "sample may not be representative."

Diagnostic categories and ROM

ROM	ND	Benign	Atypical	Suspicious	Malignant
Gupta P, Cancer Cytopathol 2021	27.5%	11.5%	66.7%	88.0%	99.6%
Vigliar E, Diagnostics 2021	50%	1.92%	58.3%	100%	100%
Torres Rivas HE, Acta Cytol 2021	27%	3%	50%	100%	100%
Caputo A, Acta Cytol 2022	46%	1.05%	28.6%	100%	99.8%
Makarenko V Cancer Cytopathol 2021	58.3%	6.4%	69.2%	96.7%	99.3%
Uzun E, Diagn Cytopathol 2022	16.6%	0.7%	88.8%	100%	100%
Ahuja S, Cytopathol 2022	9.1%	1.5%	37.5%	96.9%	98.2%



Benign Category

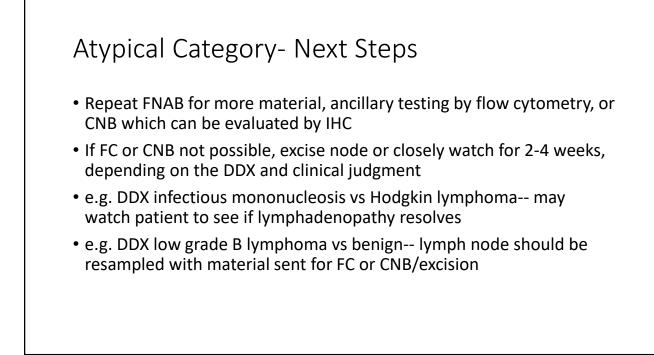
- Potentially difficult DDX:
 - Follicular hyperplasia vs follicular lymphoma
 - EBV mononucleosis vs Hodgkin lymphoma
 - Partial involvement of lymph node by low grade lymphoma
- Cytopathologist categorization of inflammatory processes as <u>Benign</u> <u>vs Atypical</u> depends on skill and practice milieu
- **If the findings raise the possibility of lymphoma, do not use Benign category- use Atypical category

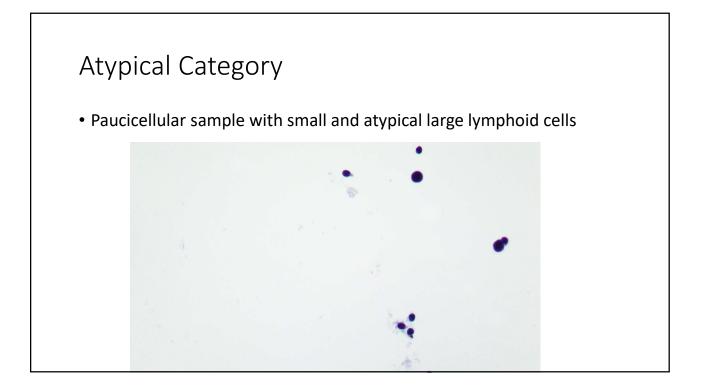
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Diagnostic categories and ROM

Atypical Category

- Predominantly supports a benign process, <u>but</u> minimal features raise possibility of malignancy
- Insufficient quantity or quality of concerning features to diagnose as benign or malignant
- ALUS: "Atypical lymphoid cells of uncertain significance" (concern for lymphoma)
- AUS: "Atypia of uncertain significance" (concern for non-lymphoid neoplasm)
- Report should describe the specific atypical features seen, and DDX raised
- The Atypical category allows Benign category to have high negative predictive value, but do not use as a "garbage can"







- Cyto slides with heterogeneous lymphoid population with tangiblebody macrophages, dendritic cells, germinal center fragments – suggests reactive follicular hyperplasia; no FC available
- Can diagnose Atypical (suggestive of FH), recommend repeat sampling with FC if LAD persists



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Suspicious for Malignancy Category

- Morphologic features concerning for malignancy
- Limited quantity/quality of findings precludes definitive Malignant dx
- High positive predictive value for Malignancy
- Includes lymphoid and non-lymphoid neoplasms

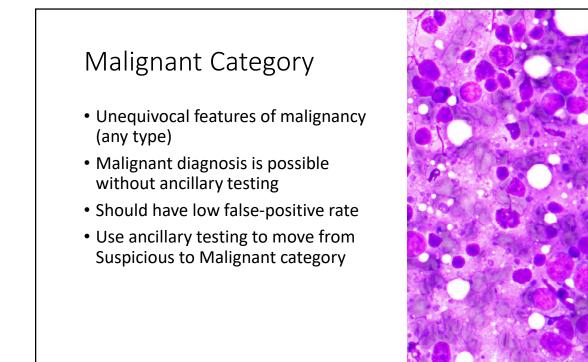
Suspicious for Malignancy

Suspicious for Malignancy Category

- Report describes the suspicious features, and provides DDX
- Utilize ancillary testing (FC, cell block with ICC) to try to move diagnosis to Malignant category
- Additional management required. Repeat sampling by FNAB or CNB +ancill. Excised if ancillary testing limited/not available.

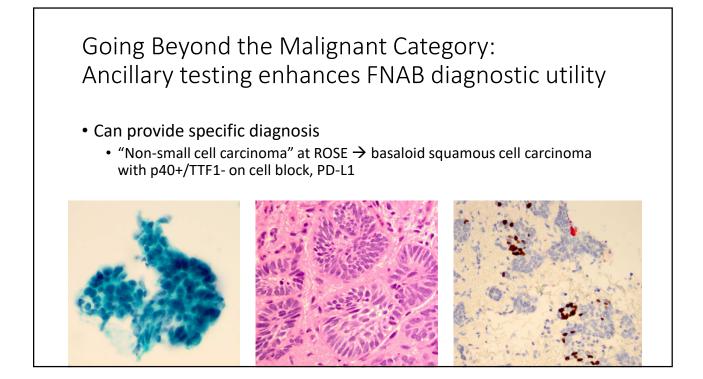
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Diagnostic categories and ROM

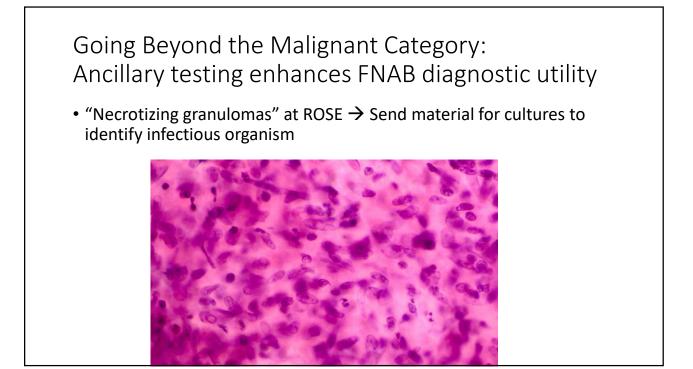
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Going Beyond the Malignant Category: Ancillary testing enhances FNAB diagnostic utility

• "Lymphocytes" at ROSE \rightarrow Mantle cell lymphoma with FC, cell block +stains



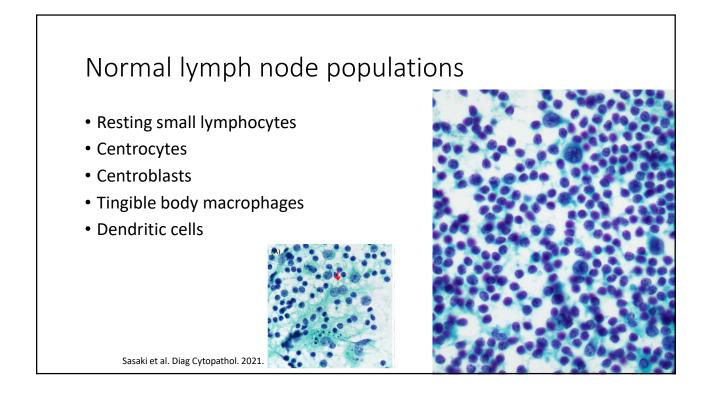


<section-header> FNA Diagnostic Utility is enhanced by: High quality FNAB smear preparations (learn and teach good technique!) ROSE decreases ND rates supports appropriate triage for ancillaries provides preliminary diagnoses Routine use of FC and Stains to confirm diagnoses FNAB can triage management for the patient (even if specific diagnosis cannot be made)

Lymph Node FNAB Sampling and Specimen Preparation

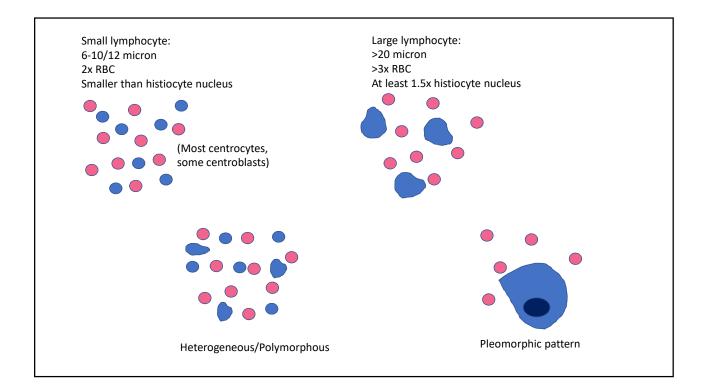
- Split sample and create multiple direct smears
- Make air-dried (Diff-Quik) and alcohol-fixed (Pap) slides, they are complementary:
 - air-dried smears better for background material and cytoplasm quality
 - alcohol-fixed better for nuclear detail
- ROSE is possible with both fixation methods
- If ROSE suggests infection, may reserve few air-dried smears for micro stains (GMS, Gram etc)
- Take additional DEDICATED passes for cultures, flow cytometry, cell block, molecular (based on ROSE)

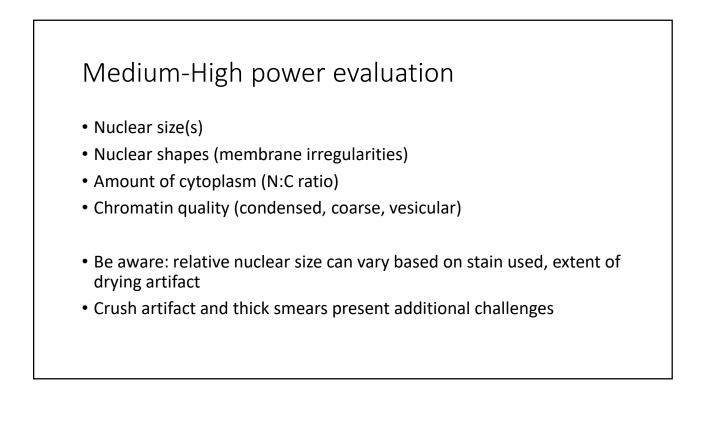
Morphologic Approach

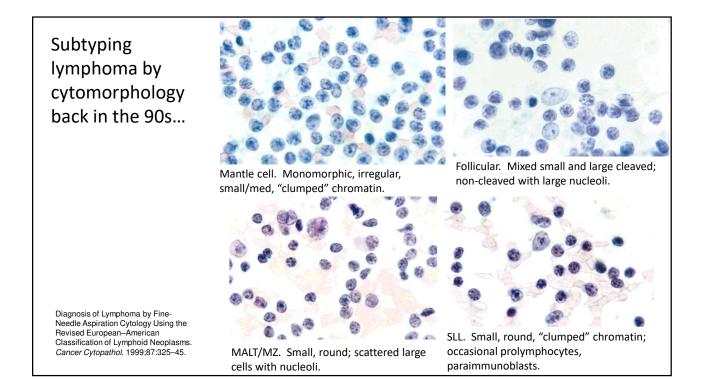


Low-power morphologic assessment

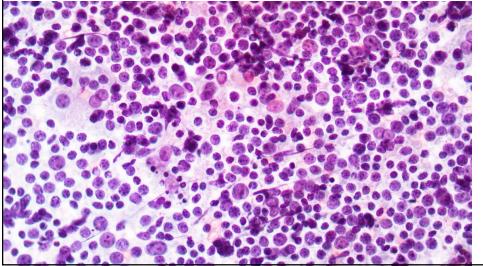
- 1. Are there cells to evaluate?
- 2. Are the cells lymphoid or non-lymphoid?
- 3. If lymphoid, identify the pattern:
 - Heterogeneous/polymorphous
 - Monotonous, small
 - Monotonous, medium
 - Monotonous, large
 - Pleomorphic







Case 1. Axillary node, 45yo man, HIV+



What pattern?

Heterogeneous/polymorphous

Monotonous, small Monotonous, medium Monotonous, large

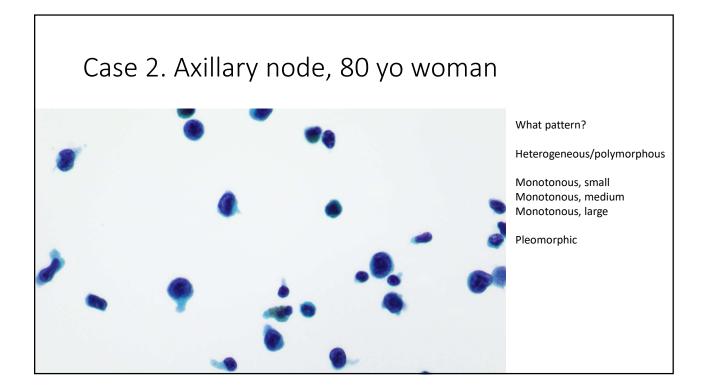
Pleomorphic

Case 1. Heterogeneous/Polymorphous pattern

DDX:

- Reactive lymphoid hyperplasia (non-specific)
- Infection
- Small B cell lymphomas (e.g. follicular, mantle, marginal zone)

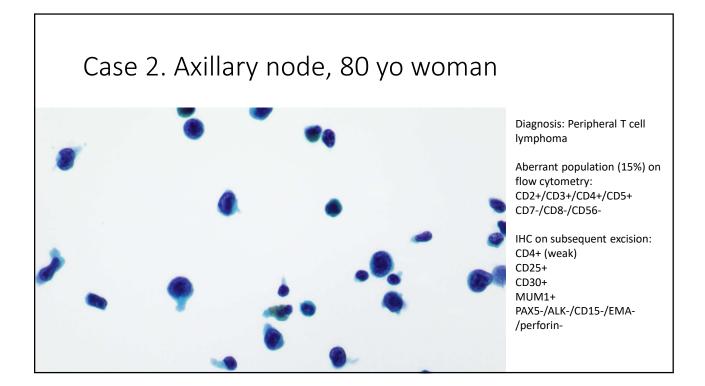
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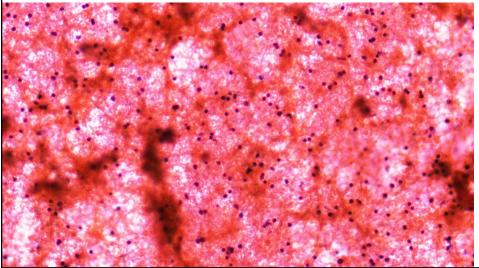
Case 2. Monotonous, Large cell pattern

DDX:

- DLBCL
- Grade 3 follicular lymphoma
- Transformation of low grade B lymphomas
- T cell lymphomas
- Blastoid mantle cell lymphoma
- Non-lymphoid metastases



Case 3. Retroperitoneum, 62 yo man



What pattern?

Heterogeneous/polymorphous

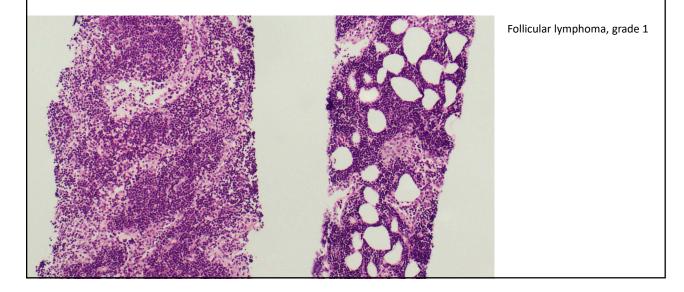
Monotonous, small Monotonous, medium Monotonous, large

Pleomorphic

Case 3. Monotonous, small cell pattern

- Reactive lymphoid hyperplasia
- Small B cell lymphomas (e.g. SLL/CLL, follicular, mantle cell, marginal zone)
- Small cell/neuroendocrine metastases

Case 3. Retroperitoneum, 62 yo man

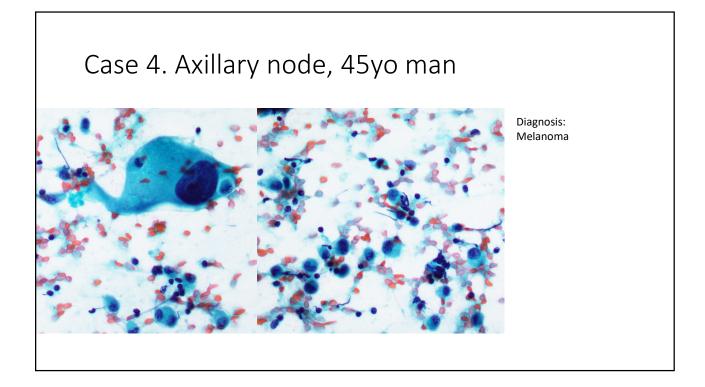


Case 4. Axillary node, 45yo man What pattern? Hetrogeneous/polymorphous Nonotonous, small Monotonous, large Pemorphic Demorphic

Case 4. Pleomorphic pattern

DDX:

- Hodgkin lymphoma
- Anaplastic large cell lymphoma
- DLBCL
- Transformation of low grade B lymphoma
- T cell lymphomas
- Metastatic malignancies

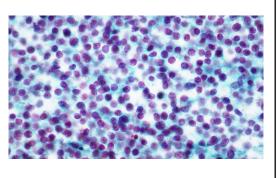


Tailor IHC panel to morphologic DDX

- Small B-cell lymphomas
- Hodgkin lymphoma
- Large cell lymphomas
- T-cell lymphomas

"Small B-cell lymphomas" suggested panel

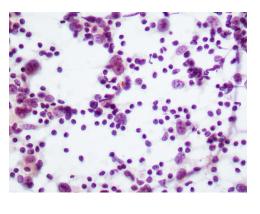
CD3	CLL/SLL
CD20	Follicular lymphoma
CD5	Mantle cell lymphoma
Cyclin D1	Marginal zone lymphoma
Sox11	Lymphoplasmacytic lymphoma
BCL2	
BCL6	
Ki67	
LEF1	



Mantle cell lymphoma

"Hodgkin" suggested panel

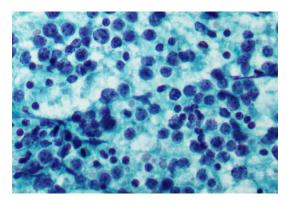
CD3	Classic Hodgkin lymphoma
CD20	NLPHL
CD30	T cell/histiocyte-rich large B cell lymphoma
CD15	Reactive node (e.g. with many immunoblasts)
EBER	
PAX5	



Hodgkin lymphoma

"Large cell lymphomas" suggested panel

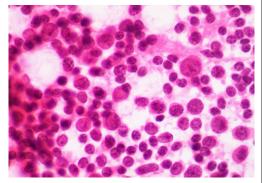
CD3, CD5	DLBCL
CD10, CD20	High grade B cell lymphoma
Cyclin D1, MUM1	Burkitt lymphoma
BCL2, BCL6	
CD30, EBER	
Ki67, C-MYC, p53	



DLBCL

"T cell lymphomas" suggested panel

CD2, CD3, CD4, CD5, CD7, CD8	Peripheral T cell lymphoma, NOS
CD25, CD30, CD56	ALCL (+/- ALK subtypes)
CD10, BCL2, BCL6	Reactive hyperplasia
CD21/CD23	Nodal T-follicular helper cell lymphomas (e.g. AITL)
Ki67, ALK, EBER, Ki67, PD-1	
Perforin, granzyme B	
TCRs (gamma, delta)	



Peripheral T cell lymphoma, NOS

