

Current Concepts in Hematopathology

Nodal Small B-cell Lymphomas

Challenging Situations and Potential Pitfalls

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SMALL B-CELL LYMPHOMAS

Classification

CD5+

- Chronic lymphocytic leukemia/small lymphocytic lymphoma
 - Monoclonal B-cell lymphocytosis
- Mantle cell lymphoma
 - Leukemic non-nodal mantle cell lymphoma
 - In situ mantle cell neoplasia

CD10+

- Follicular lymphoma
 - Testicular follicular lymphoma
 - In situ follicular neoplasia
 - Duodenal-type follicular lymphoma
- Pediatric-type follicular lymphoma
- Large B-cell lymphoma with IRF4 rearrangement
- Primary cutaneous follicle center lymphoma

CD5-/CD10-

- Lymphoplasmacytic lymphoma
- Hairy cell leukemia
- Splenic B-cell lymphoma/leukemia, unclassifiable
 - Splenic diffuse red pulp B-cell lymphoma
 - Hairy cell leukemia, variant
- Splenic marginal zone lymphoma
- Extranodal marginal zone lymphoma of mucosa-associated lymphoid tissue (MALT lymphoma)
- Nodal marginal zone lymphoma
 - Pediatric nodal marginal zone lymphoma

CD5-/+

- B-cell prolymphocytic leukemia

CHRONIC LYMPHOCYTIC LEUKEMIA/SMALL LYMPHOCYTIC LYMPHOMA

Gray areas/caveats

Monoclonal B-cell lymphocytosis (MBL)

- circulating clonal B cells $<5 \times 10^9/L$
- no documented lymphadenopathy, organomegaly, other extramedullary disease
- patients with MBL can have bone marrow involvement (interstitial, non-paratrabeular lymphoid aggregates, diffuse) – ranges, but on average accounts for ~20% of the cellularity
- difference in definitions:
 - *International Workshop on CLL* (iwCLL)*: “the presence of a cytopenia caused by a typical marrow infiltrate establishes the diagnosis of CLL regardless of the number of peripheral blood B lymphocytes or of the lymph node involvement”
 - WHO: “the [4th ed] revision will eliminate the option to diagnose CLL with $<5 \times 10^9/L$ peripheral blood CLL cells in the absence of extramedullary disease, even if there are cytopenias or disease-related symptoms”

Limited nodal involvement by CLL-like cells

- sometimes this is an incidental flow finding in lymph nodes taken out for other reasons (subtle by morphology/IHC) → recommend PB flow cytometry evaluation to exclude circulating component at $\geq 5 \times 10^9/L$
- if circulating CLL-like count is $<5 \times 10^9/L$, probably best considered “nodal MBL” (*not* SLL) if: architecture not effaced; proliferation centers not identified; lymph nodes are $<1.5\text{cm}$ by imaging

Hallek M et al. *Blood*. 2018;131(25):2745-2760.
Ganapathi KA et al. *Haematologica*. 2014; 99(9). Gibson SE et al. *Haematologica*. 2011;96(8):1144-1152.
Strati P, Shanafelt TD. *Blood*. July 23 2015;126(4). Randen U et al. *Am J Clin Pathol*. 2013;139:390-395.

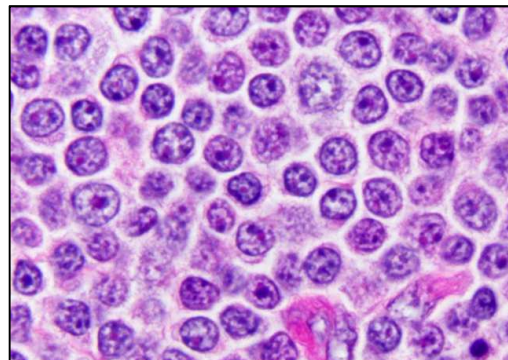
CHRONIC LYMPHOCYTIC LEUKEMIA/SMALL LYMPHOCYTIC LYMPHOMA

Proliferation centers



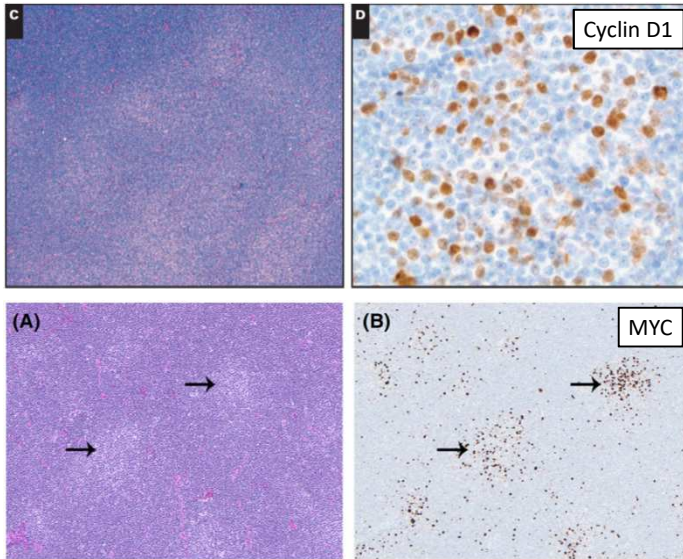
Lymph node:

- diffuse effacement; occasionally vaguely nodular
- proliferation centers: small lymphocytes, prolymphocytes and paraimmunoblasts



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Proliferation centers – potential pitfalls



- proliferation centers in up to 30% of cases of CLL/SLL can have cyclin D1 expression
- no t(11;14)
- no SOX11 expression by IHC
- does not = mantle cell lymphoma
- proliferation centers in the majority of cases of CLL/SLL have at least subset MYC expression by IHC
- no MYC rearrangement by FISH; few cases with MYC hyperdiploidy by FISH
- does not = transformation to large-cell lymphoma

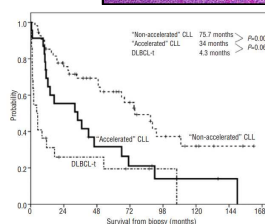
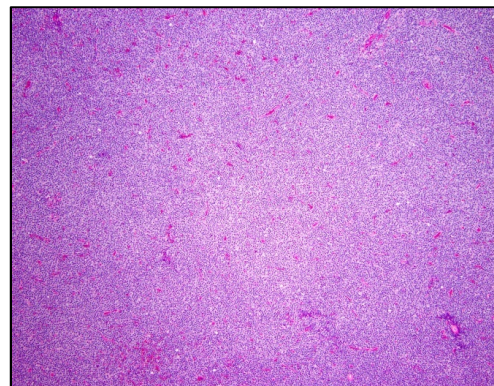
Gibson SE et al. *Br J Haematol.* 2016;175, 161–175.
Gradowski JF et al. *Am J Clin Pathol.* 2012;138:132–139.

CHRONIC LYMPHOCYTIC LEUKEMIA/SMALL LYMPHOCYTIC LYMPHOMA

Proliferation centers - confluent

- proliferation centers broader than 20x field or becoming confluent and/or Ki67 >40% or mitoses >2.4/PC
- association with deletion in 17p13 or trisomy 12
- has been referred to as “accelerated CLL”
- not an official category in the current WHO
- challenging “gray zone” histologically
- distinct from DLBCL (often requires excisional biopsy)
 - DLBCL = confluent sheets of large B cells with a nuclear size equal to or exceeding that of normal macrophage nuclei or more than twice the size of a normal lymphocyte
- clinical trial study showed that only 33 of 40 (82.5%) cases submitted as Richter transformation were consistent with RT following expert central review
 - large proliferation centers
 - variably confluent and serpiginous proliferation centers
 - high proliferation index (sometimes thick section or associated normal bone marrow)
- comment upon in report given association with more aggressive clinical course
 - note that outcomes studies were before the current therapy era

Confluent

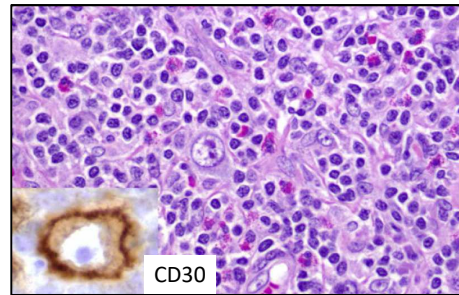
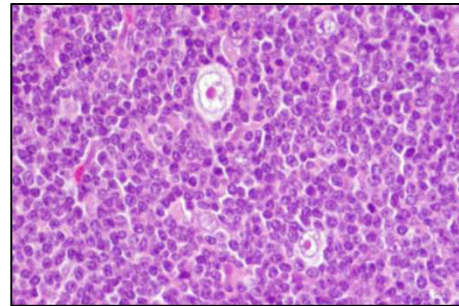


Soilleux EJ et al. *Histopathology.* 2016;69, 1066–76.
Gine E et al. *Haematologica.* 2010;95(9):1526–33.
Ciccone M et al. *Leukemia.* 2012;26:499–508.
Oscier D et al. *Br J Haematology.* 2016;174, 767–775.
Chabot-Richards D et al. *Chp 14. Hematopathology 2nd ed.* 2016.

CHRONIC LYMPHOCYTIC LEUKEMIA/SMALL LYMPHOCYTIC LYMPHOMA

Hodgkin/Reed-Sternberg (HRS) cells

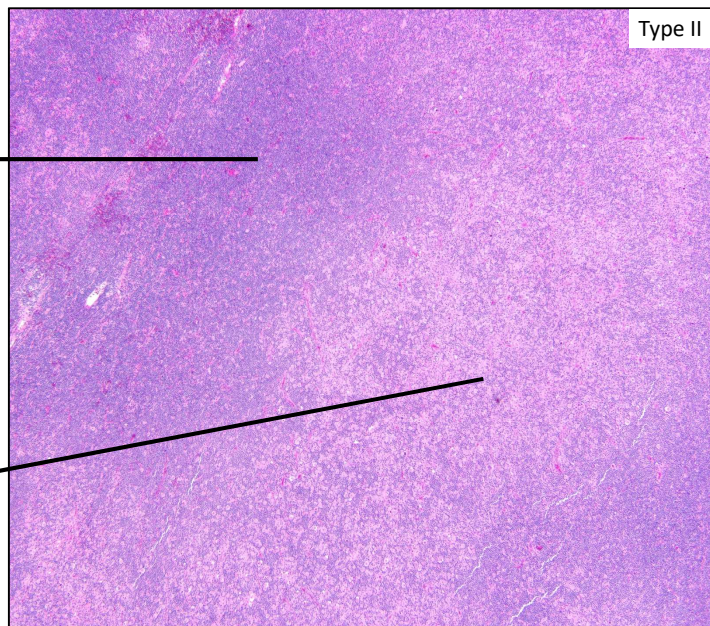
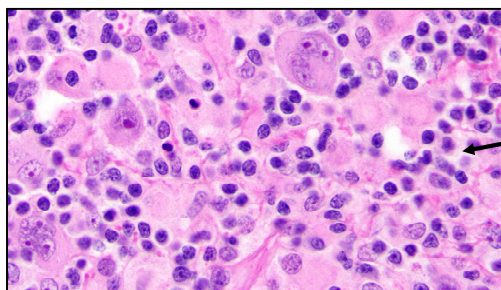
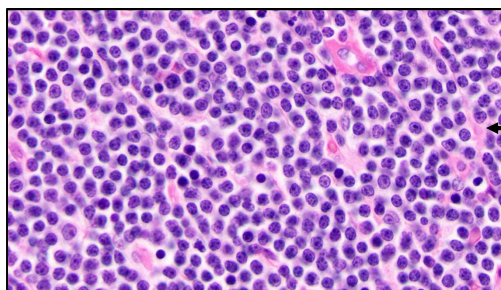
- “Type I” pattern, or CLL/SLL with HRS-like cells (CLL-HRS)
 - HRS cells sparsely distributed amongst CLL cells
 - median 4 HRS cells/high-powered field; often with T-cell rosette
 - no evidence of inflammatory background typical of classic Hodgkin lymphoma
 - per 2017 WHO, does not fulfill criteria for CHL/not “Richter transformation”
 - most EBV+
 - may or may not be clonally related to CLL
- “Type II” pattern, or Richter transformation as CHL (RT-CHL)
 - requires diagnostic RS cells in an inflammatory background typical of CHL per WHO
 - most EBV+
 - ~50% related to the CLL clone; clonally-unrelated cases associated with unmutated CLL
 - uncommon type of Richter transformation (~5%)
 - inferior prognosis compared to *de novo* HL



King RL et al. *Blood Cancer Journal*. 2022;12:18.
 Xiao W et al. *Hum Pathol*. 2016 September;55: 108–116.
 Chabot-Richards D et al. *Chp 14. Hematopathology 2nd ed.* 2016.

CHRONIC LYMPHOCYTIC LEUKEMIA/SMALL LYMPHOCYTIC LYMPHOMA

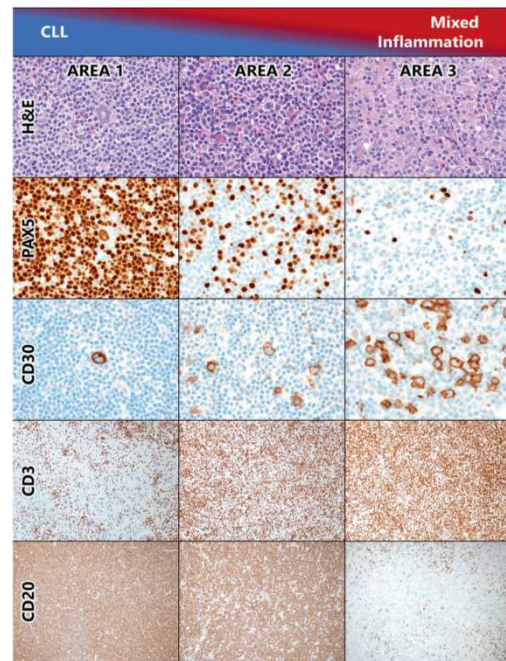
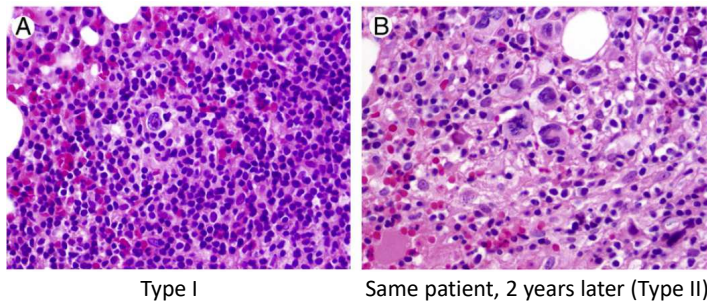
Hodgkin/Reed-Sternberg (HRS) cells



CHRONIC LYMPHOCYTIC LEUKEMIA/SMALL LYMPHOCYTIC LYMPHOMA

Hodgkin/Reed-Sternberg (HRS) cells

- however, other cases can present as a histologic continuum or can progress over time



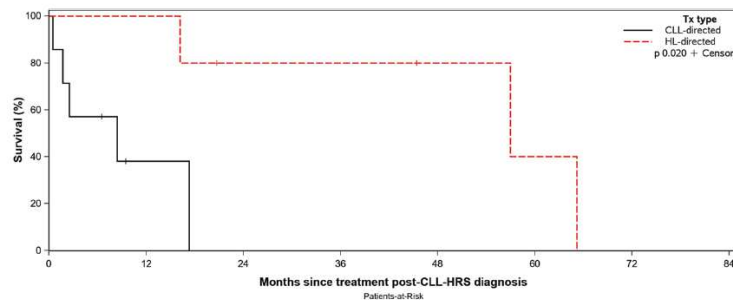
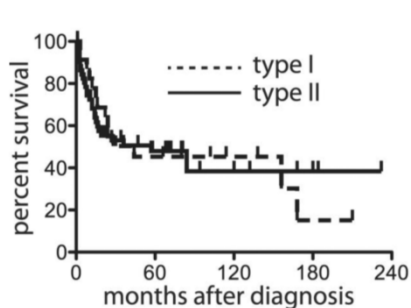
King RL et al. *Blood Cancer Journal*. 2022;12:18.

Xiao W et al. *Hum Pathol*. 2016 September; 55: 108–116.

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Hodgkin/Reed-Sternberg (HRS) cells

- Do we need to make a distinction between CLL-HRS and RT-CHL?



(retrospective study spanning 3 decades)

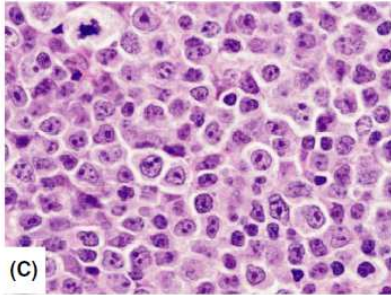
King RL et al. *Blood Cancer Journal*. 2022;12:18.

Xiao W et al. *Hum Pathol*. 2016 September; 55: 108–116.

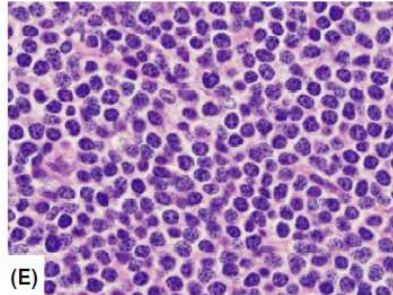
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Pseudo-Richter transformation

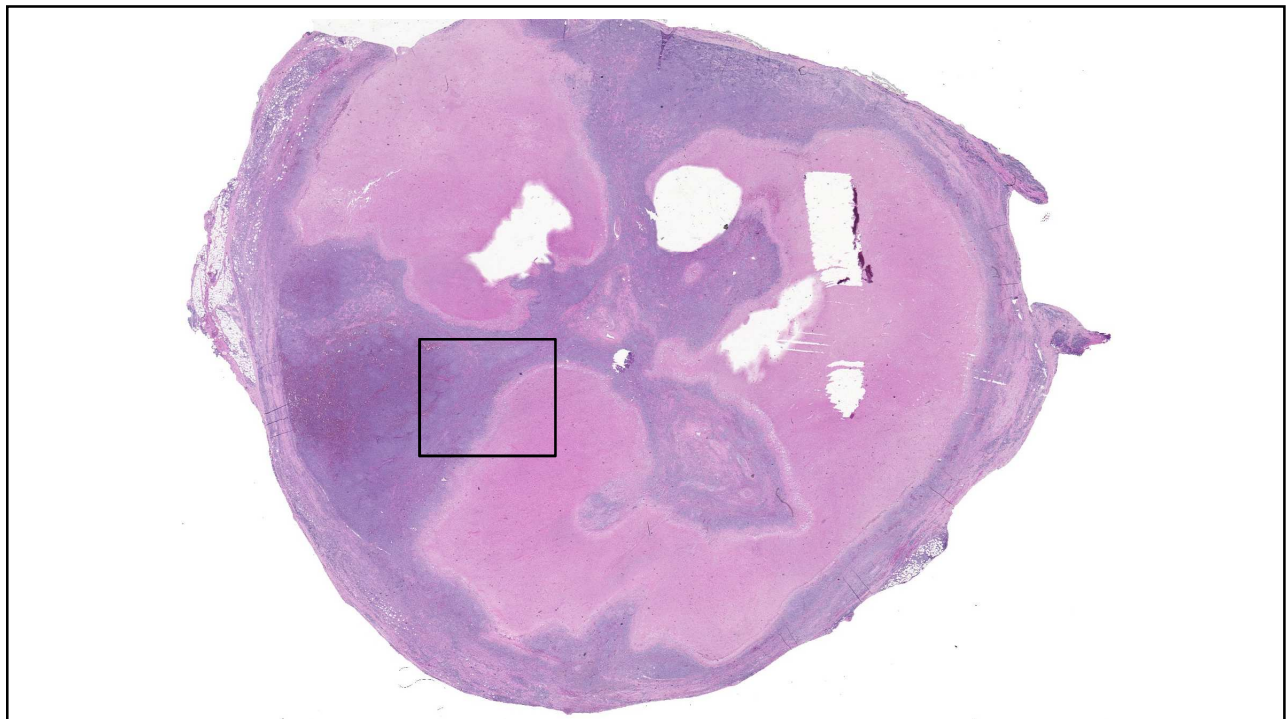
- RT occurs at a similar rate with novel agents like ibrutinib (BTK inhibitor) as traditional chemotherapy agents, although rate is lower if front-line
- If RT occurs, it is usually an early event (within the first year) and clonally related to CLL
- recent report of 5 patients with CLL/SLL receiving ibrutinib who underwent brief interruption of therapy due to upcoming surgery → developed lymphadenopathy, lymphocytosis or progressive cytopenias
- biopsies (LN or bone marrow) revealed findings morphologically consistent with RT
- histologic findings resolved upon resumption of therapy

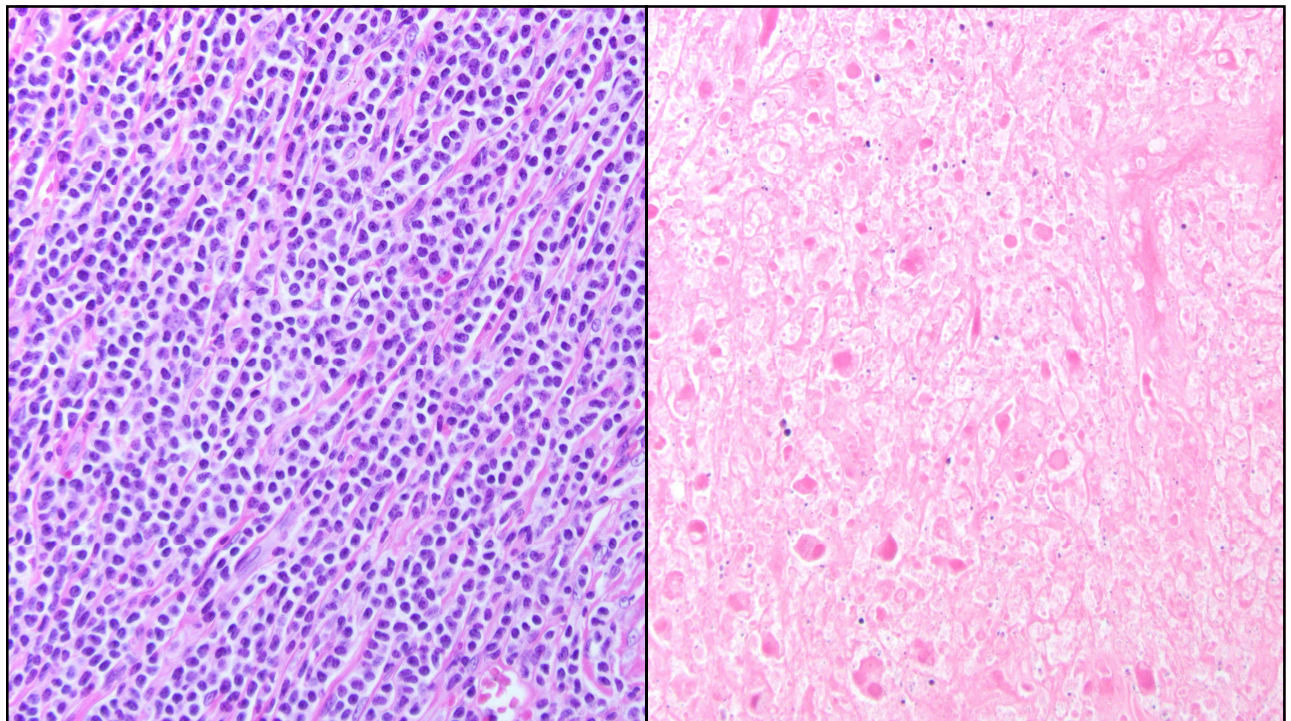
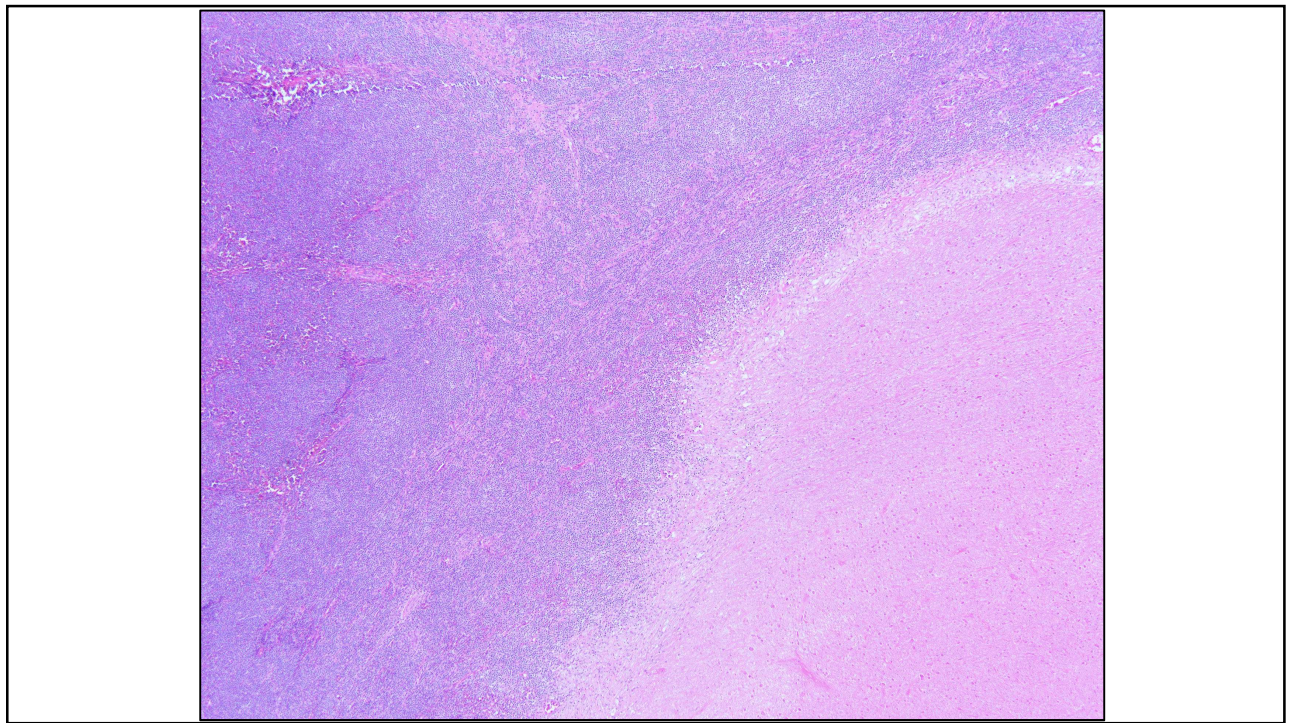


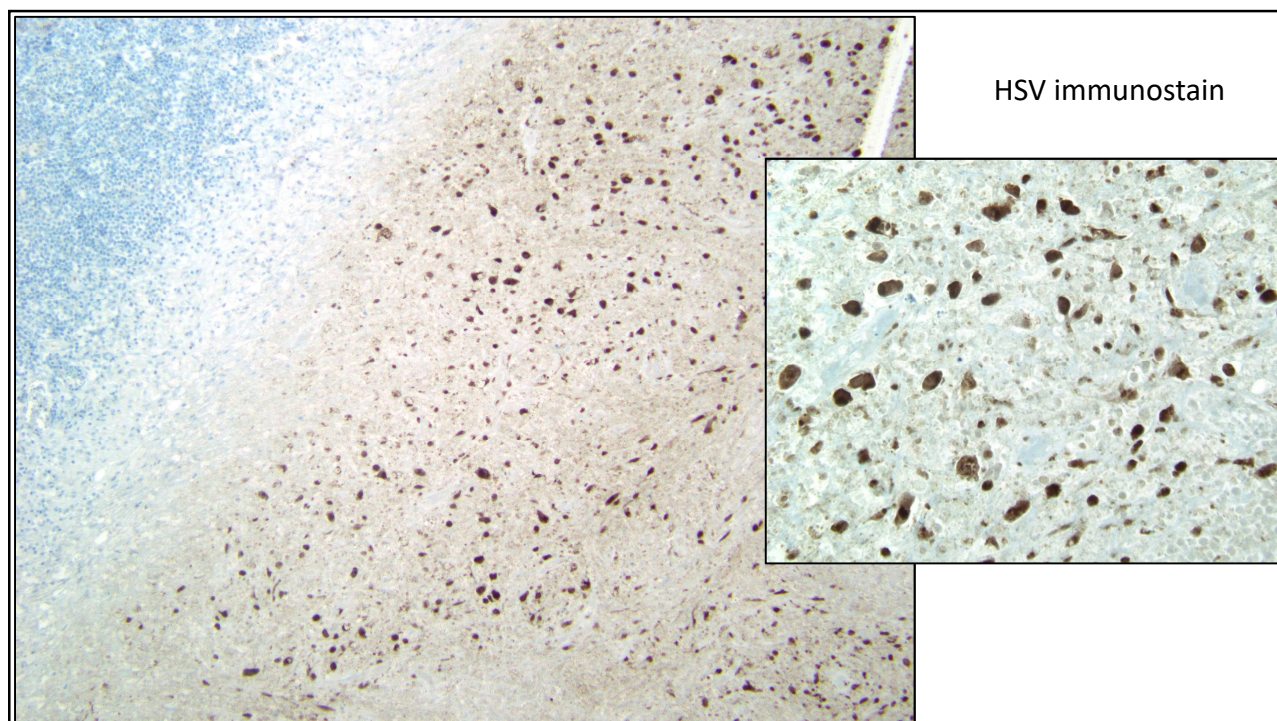
Biopsy 7-13 days into therapy interruption



Biopsy following resumption of therapy





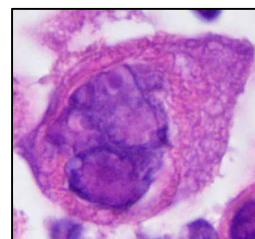


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Necrosis – potential pitfall

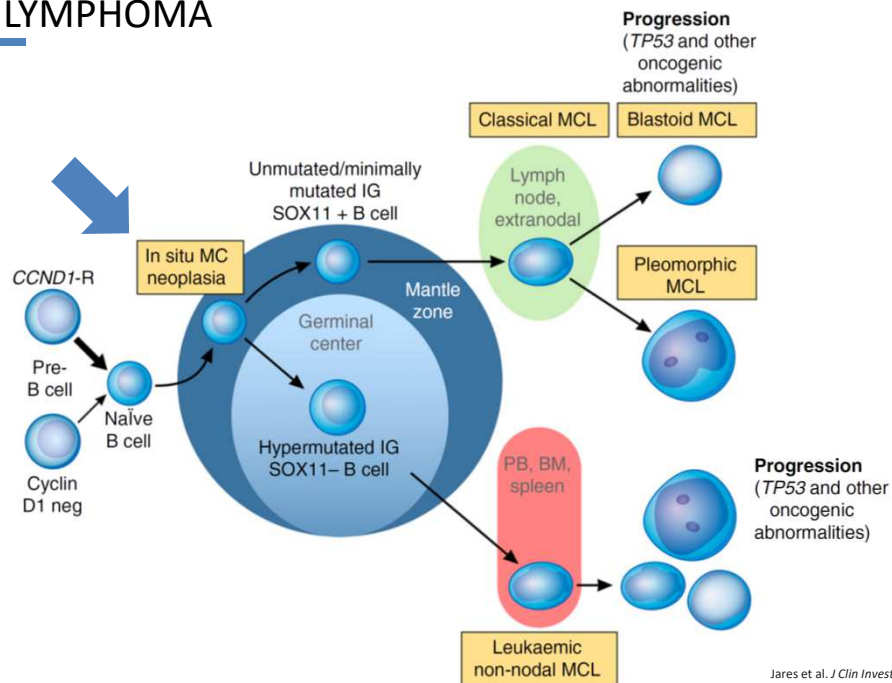
Herpes simplex virus (HSV) lymphadenitis

- well-circumscribed areas of necrosis with nuclear dust/debris, eosinophilic ghost cells, viral inclusions, neutrophils
- HSV-infected cells can be mononuclear or multinucleated, and show eosinophilic, “ground-glass” intranuclear inclusions
- can infect any cell type
- reactivation of HSV associated with lymphadenopathy is particularly seen in patients with CLL/SLL (?secondary to immunosuppression)
- can also be seen in other hematologic malignancies
- *do not interpret necrosis as unequivocal evidence of Richter transformation – may be HSV!*



MANTLE CELL LYMPHOMA

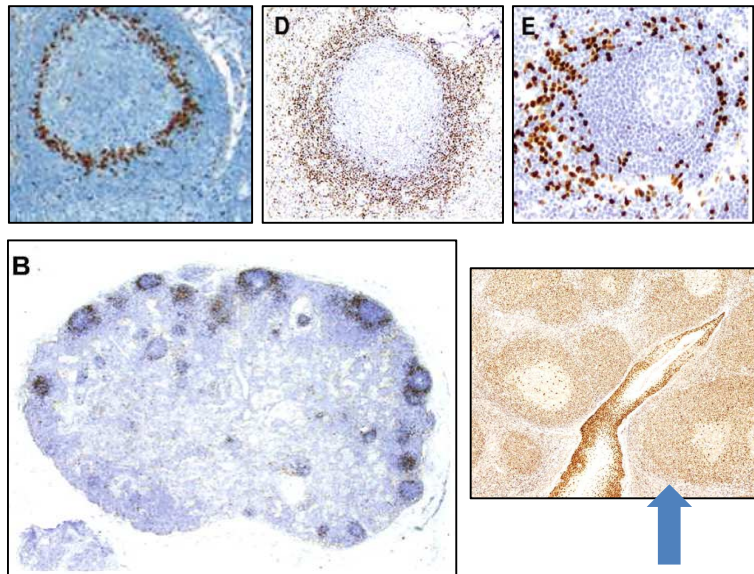
New paradigm



MANTLE CELL LYMPHOMA

In situ mantle cell neoplasia

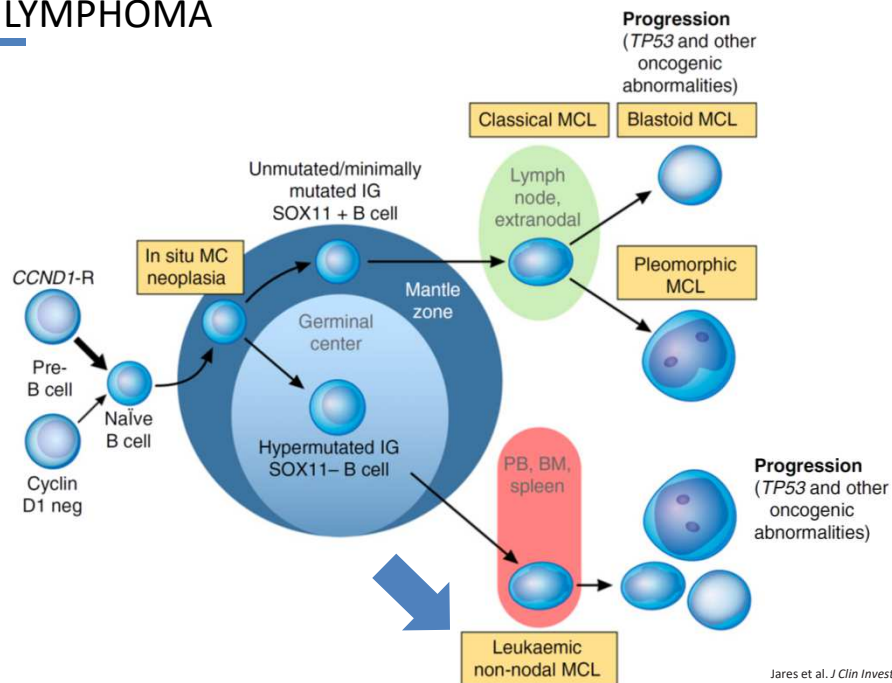
- non-expanded mantle zones
- cyclin D1⁺ cells in inner, outer or entire mantle zone
- no other features in the lymph node of MCL
- often found incidentally, particularly in association with a true lymphoma
- t(11;14) present; SOX11+/-
- low rate of progression
- the term “neoplasia” emphasizes need for conservative clinical approach → not equal to MCL/typically does not show progression to MCL



Caveat! Must be distinguished from overt MCL with a mantle zone growth pattern = expanded mantle zones

MANTLE CELL LYMPHOMA

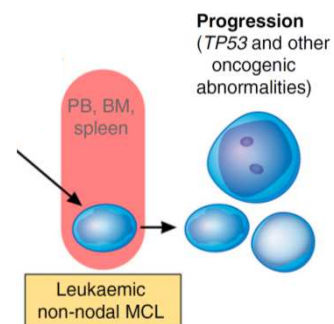
New paradigm



MANTLE CELL LYMPHOMA

Leukemic non-nodal MCL

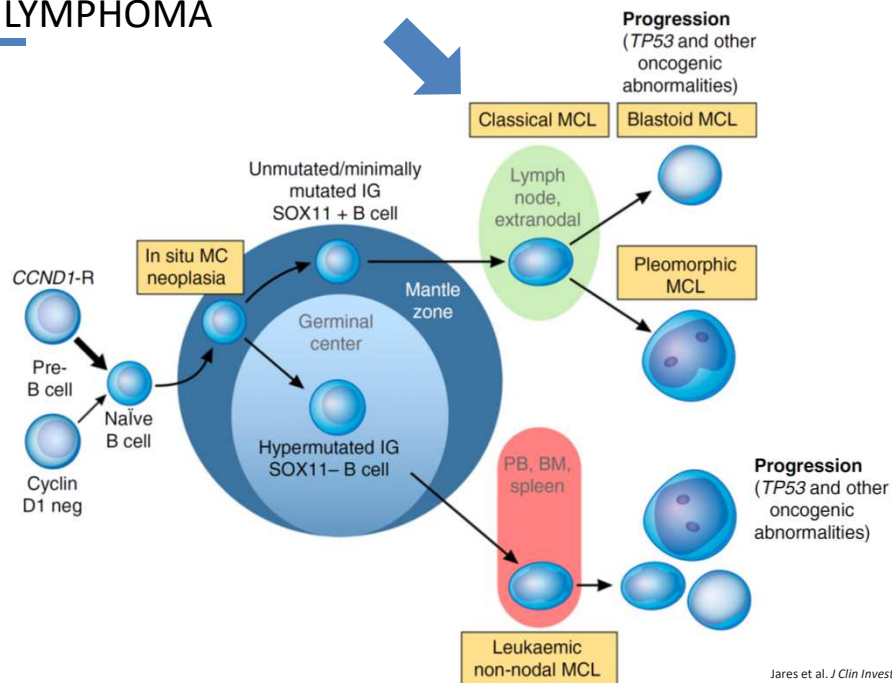
- peripheral blood, bone marrow and splenic involvement without significant adenopathy
- SOX11-negative
- may be CD5-negative, CD200+
- morphologically may resemble CLL/SLL
- somatic hypermutation
- t(11;14) present; few if any additional karyotypic abnormalities
 - ATM mutation/11q22 deletion rare to absent
- more indolent course/better overall survival
- acquisition of TP53 mutation or other oncogenic gene abnormalities may result in more aggressive behavior



Consider this entity in evaluation of leukemic lymphomas – always perform cytogenetic evaluation for t(11;14) in this setting

MANTLE CELL LYMPHOMA

New paradigm

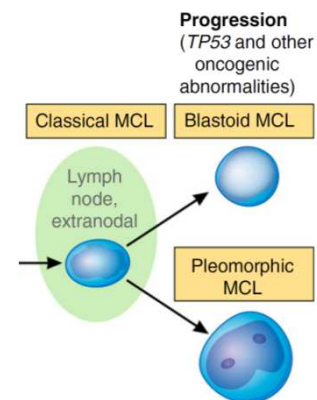


Jares et al. *J Clin Invest.* 2012;122:3416-3412.
Swerdlow S et al. *Blood.* 2016;127(20):2375-2390.

MANTLE CELL LYMPHOMA

"Classical" MCL

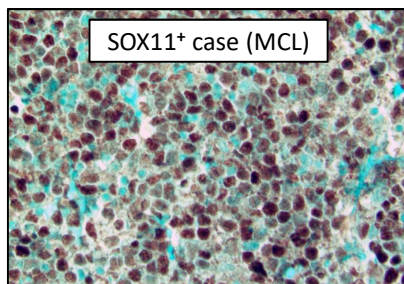
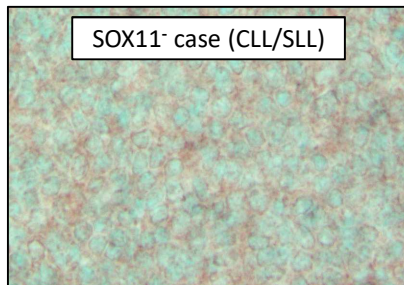
- involves lymph nodes and extranodal sites
- SOX11+, CD200-negative
- no evidence of transit through germinal center (unmutated)
- genetically unstable and acquires secondary driver mutations in cell cycle regulatory genes, DNA damage response pathway, cell survival mechanisms
 - ATM mutation/11q22 deletion common
- clinically aggressive course
- may progress to blastoid or pleomorphic MCL



Swerdlow S et al. *Blood.* 2016;127(20):2375-2390.

MANTLE CELL LYMPHOMA

SOX11



The choice of SOX11 antibody matters!

TABLE 1. IHC SOX11 Expression in a Series of Cyclin D1-positive or Cyclin D1-negative Lymphomas, Using the Mouse Monoclonal anti-SOX11^{180Q-38}, Anti-SOX11¹⁴³, and the Goat Polyclonal Anti-SOX11¹⁶⁻¹⁷³⁴⁷ Antibody

CCND1	Diagnosis	Mouse Monoclonal Anti-SOX11 ^{180Q-38} SOX11 Positive/Total Cases (%) (n = 205)	Mouse Monoclonal Anti-SOX11 ¹⁴³ SOX11 Positive/Total Cases (%) (n = 209)	Goat Polyclonal Anti-SOX11 ¹⁶⁻¹⁷³⁴⁷ SOX11 Positive/Total Cases (%) (n = 177)
Negative	B-LBL	0/4	2/4 (50)	0/4
	T-LBL	0/4	4/4 (100)	1/3 (33)
	CLL	0/9	0/9	0/9
	HCL	0/3	0/3	1/3 (33)
	PCM/Plasmacytoma	0/4	0/4	0/3
	MZL	0/10	0/10	0/10
	FL	0/8	0/8	0/1
	MCL	9/9 (100)	9/9 (100)	9/9 (100)
	DLBCL	0/43	2/43 (5)	1/35 (3)
	PBL	0/9	0/9	0/7
	PEL	0/2	0/2	0/2
	BL	0/32	22/32 (69)	6/32 (19)
	BCLL	0/5	0/5	1/4 (25)
	T-PLL	0/2	1/2 (50)	0/2
	Extranodal NK/T-cell lymphoma	0/1	0/1	0/1
	EATL II	0/1	0/1	0/1
	HSTL	0/3	0/3	0/3
	PTCL, NOS	0/11	0/11	0/10
	AITL	0/6	0/6	0/4
	ALCL ALK ⁺	0/3	0/3	0/3
	ALCL ALK ⁻	0/1	0/1	0/1
Positive	NLPHL	0/1	0/1	—
	cHL	0/5	0/5	0/1
	HCL	0/2	0/2	0/2
	PCM/Plasmacytoma	0/4	0/4	0/4
	MCL	20/23 (87)	20/27 (74)	18/23 (78)

AITL indicates angioimmunoblastic T-cell lymphoma; ALCL, anaplastic large cell lymphoma; BCLL, B-cell lymphoma unclassified with features intermediate between DLBCL and BL; B-LBL, B-lymphoblastic lymphoma; cHL, classical Hodgkin lymphoma; CLL, chronic lymphocytic leukemia; EATL II, enteropathy-associated T-cell lymphoma type II; FL, follicular lymphoma; HSTL, hepatosplenic T-cell lymphoma; MZL, marginal zone lymphoma; NLPHL, nodular lymphocyte predominant Hodgkin lymphoma; PBL, plasmablastic lymphoma; PCM, plasma cell myeloma; PEL, primary effusion lymphoma; PTCL, NOS, peripheral T-cell lymphoma not otherwise specified; T-LBL, T-lymphoblastic lymphoma; T-PLL, T-cell prolymphocytic leukemia.

Soldini D et al. *Am J Surg Pathol*. 2014;38:86–93.

MANTLE CELL LYMPHOMA

Infrequent IHC and FISH variations

	Cyclin D1 IHC+ t(11;14) FISH-
Possible explanations	MCL with cryptic <i>CCND1</i> insertion into <i>IGH</i> locus (very rare) or MCL with <i>CCND1</i> fusion with <i>IGK</i> or <i>IGL</i> instead of <i>IGH</i> or other entity: plasma cell neoplasm, CLL with expanded proliferation centers, hairy cell leukemia (weak)
Next steps	NGS evaluation Break-apart probes Full IHC work-up for other entities

Sethi S et al. *Front Oncol*. 2021;11:739441.

MANTLE CELL LYMPHOMA

Infrequent IHC and FISH variations

	Cyclin D1 IHC+ t(11;14) FISH-	Cyclin D1 IHC- t(11;14) FISH+
Possible explanations	MCL with cryptic <i>CCND1</i> insertion into <i>IGH</i> locus (very rare) or MCL with <i>CCND1</i> fusion with <i>IGK</i> or <i>IGL</i> instead of <i>IGH</i> or other entity: plasma cell neoplasm, CLL with expanded proliferation centers, hairy cell leukemia (weak)	epitope not recognized by commercial antibody
Next steps	NGS evaluation Break-apart probes Full IHC work-up for other entities	SOX11 expression with classic morphologic features confirms MCL diagnosis

Sethi S et al. *Front Oncol.* 2021;11:739441.

MANTLE CELL LYMPHOMA

Infrequent IHC and FISH variations

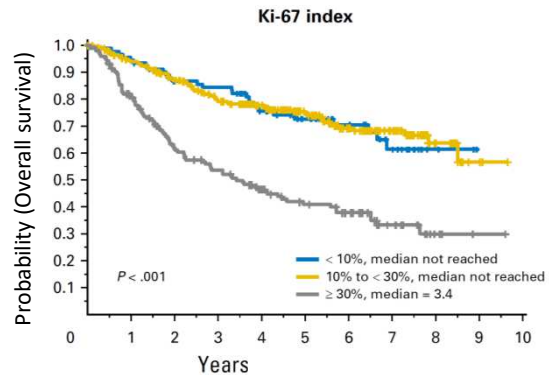
	Cyclin D1 IHC+ t(11;14) FISH-	Cyclin D1 IHC- t(11;14) FISH+	Cyclin D1 IHC- t(11;14) FISH-
Possible explanations	MCL with cryptic <i>CCND1</i> insertion into <i>IGH</i> locus (very rare) or MCL with <i>CCND1</i> fusion with <i>IGK</i> or <i>IGL</i> instead of <i>IGH</i> or other entity: plasma cell neoplasm, CLL with expanded proliferation centers, hairy cell leukemia (weak)	epitope not recognized by commercial antibody	>50% of these cases have <i>CCND2</i> rearrangements, with <i>IGK</i> , <i>IGL</i> or <i>IGH</i>
Next steps	NGS evaluation Break-apart probes Full IHC work-up for other entities	SOX11 expression with classic morphologic features confirms MCL diagnosis	SOX11 expression with classic morphologic features confirms MCL diagnosis may be positive for <i>CCND2</i> and <i>CCND3</i> by IHC, but these are non-specific (CLL, follicular lymphoma, splenic marginal zone lymphoma) NGS evaluation, FISH for <i>CCND2</i> or 3 translocation

Sethi S et al. *Front Oncol.* 2021;11:739441.

MANTLE CELL LYMPHOMA

Proliferation index

- MCL is not graded, but elevated proliferation index is associated with adverse prognosis
 - Ki67 staining: $\geq 30\%$ associated with decreased overall survival
 - # of mitoses per 15 high-powered fields (HPFs): >10
 - assessment of proliferation index should be performed in all cases of MCL

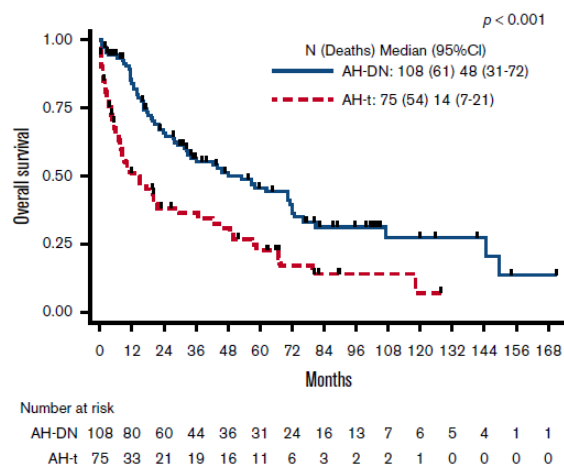


Hoster E et al. *J Clin Oncol*. 2016; 34(12):1386-1394.

MANTLE CELL LYMPHOMA

Blastoid and pleomorphic variants

- certain morphologies are associated with more aggressive behavior:
 - blastoid variant** = cells resemble lymphoblasts; often ≥ 20 -30 mitoses per 10 HPFs
 - pleomorphic variant** = large cells with pleomorphic nuclei and often prominent nucleoli
- may arise *de novo* or as transformation from MCL with classical morphology

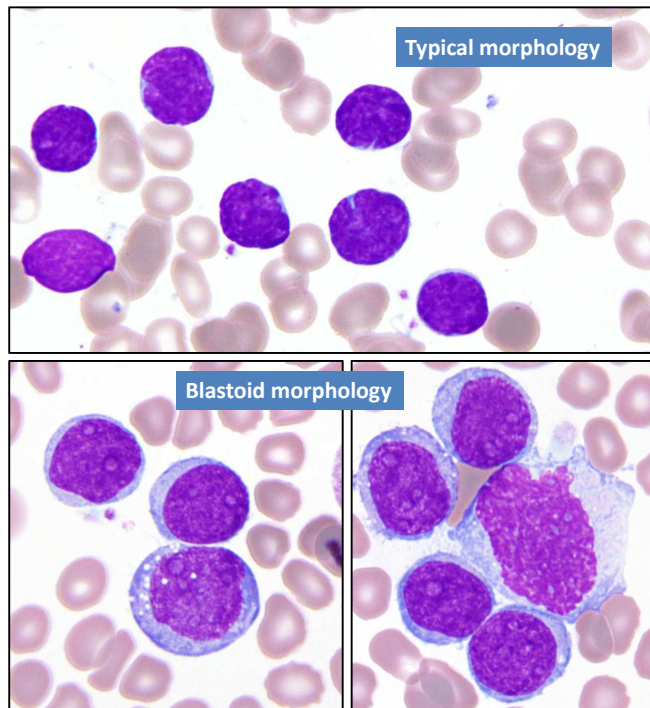


Jain P et al. *Blood Adv*. 2020;4(6): 1038-1050.
Hoster E et al. *J Clin Oncol*. 2016;34(12):1386-1394.

MANTLE CELL LYMPHOMA

Blastoid and pleomorphic variants

- morphology can be deceiving, particularly *de novo* cases
- recommend cyclin D1 in routine work-up of DLBCL and B-cell neoplasms with immature cytologic features



MANTLE CELL LYMPHOMA

TP53

- numerous recurrent gene mutations (including *ATM* and *TP53*) and chromosomal gains and losses have been described in MCL
- *TP53* gene aberrations (mutation or 17p deletion) in MCL → aggressive disease features such as blastoid/pleomorphic histology, high Ki-67, complex karyotype, high-risk MIPI (MCL International Prognostic Index) → predict poor outcomes

NCCN Guidelines B-cell lymphomas Version 2.2022

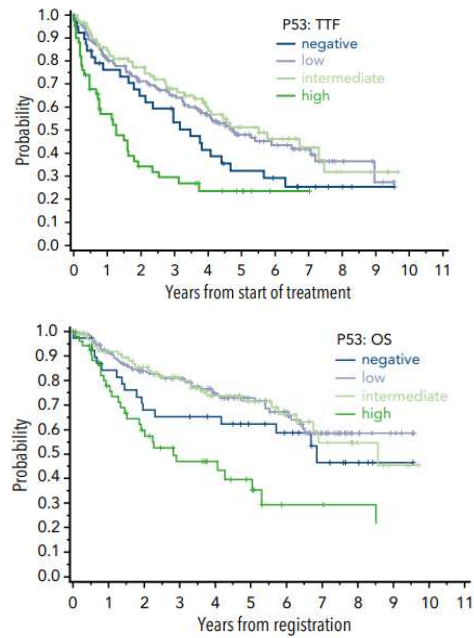
ESSENTIAL:

- Adequate immunophenotyping to establish diagnosis^a
 - IHC panel: CD20, CD3, CD5, cyclin D1, CD10, CD21, CD23, BCL2, BCL6, SOX11, Ki-67^b with or without
 - Cell surface marker analysis by flow cytometry with peripheral blood and/or biopsy specimen: kappa/lambda, CD19, CD20, CD5, CD23, CD10, CD200
- *TP53* sequencing^c

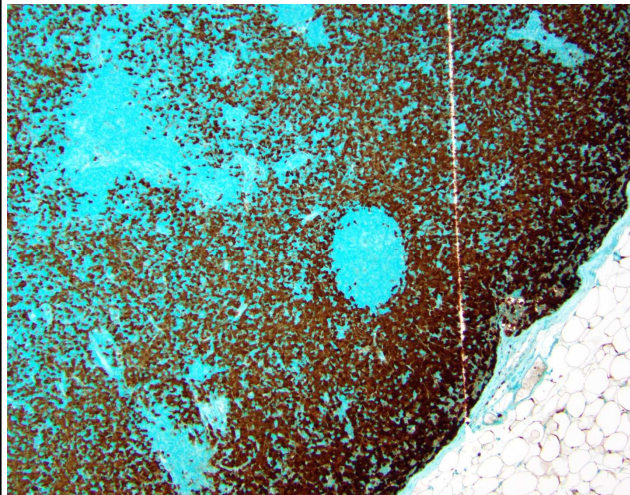
MANTLE CELL LYMPHOMA

To p53 IHC or not to p53 IHC

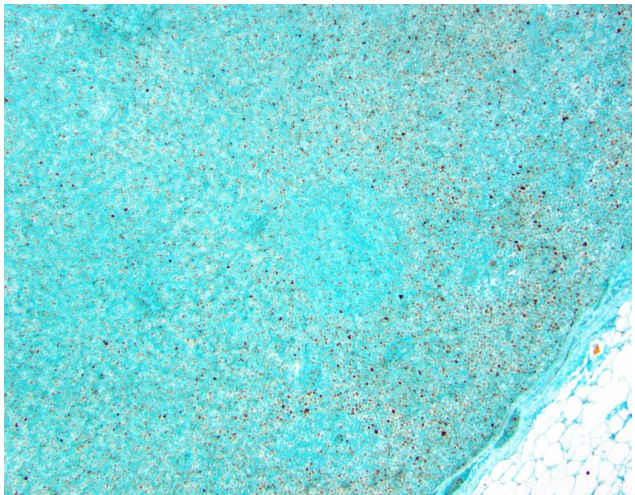
- Aukema et al *Blood* 2018: patients with high p53 expression by IHC (>50% positive lymphoma cells) had a shorter time to treatment failure and poor OS independent of both MIPI score and Ki-67 index
 - negative (0% positive lymphoma cells)
 - low (1%-10% positive lymphoma cells)
 - intermediate (10%-50% positive lymphoma cells)
 - high (>50% positive lymphoma cells)
- our clinicians have asked us to perform p53 IHC on mantle cell cases for prognostic information → affects management decisions, including re: transplant



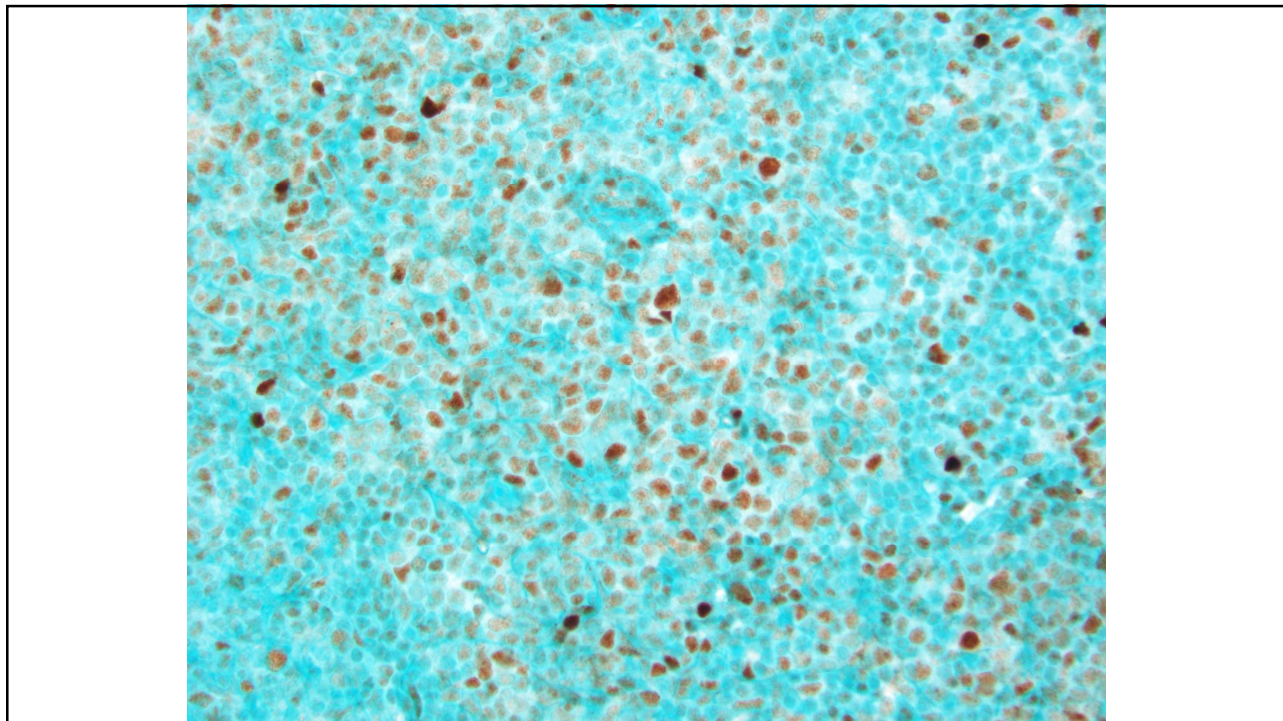
Aukema SM et al. *Blood*. 2018;131(4):417-420.



Cyclin D1



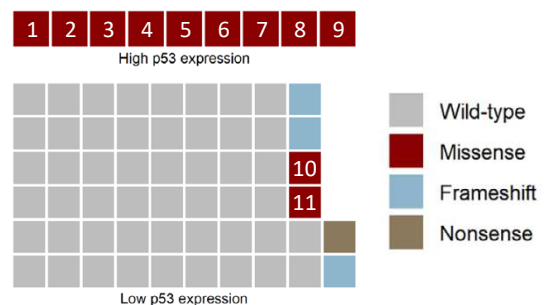
p53



MANTLE CELL LYMPHOMA

To p53 IHC or not to p53 IHC

- How does IHC correlate with mutation status?
 - p53 is overexpressed in the setting of missense mutations (due to protein accumulation)
 - Rodrigues et al *Br J Haematol* 2020:
 - >30% dark brown nuclei = p53 overexpression (faint staining was not considered)
 - p53 overexpression detected in 9 of 11 cases with p53 missense mutations



NCCN Guidelines B-cell lymphomas Version 2.2022

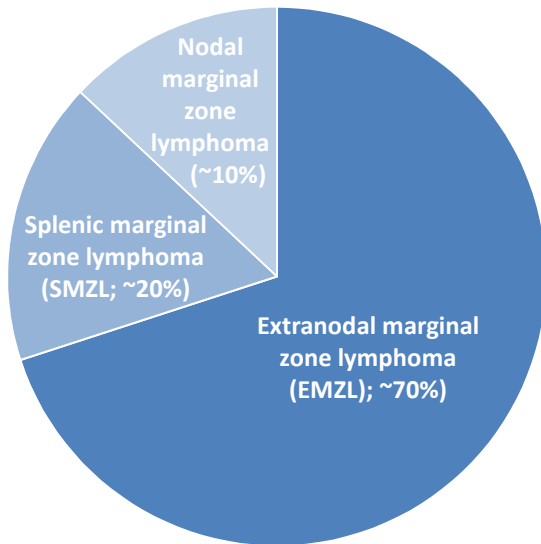
ESSENTIAL:

- Adequate immunophenotyping to establish diagnosis^a
 - IHC panel: CD20, CD3, CD5, cyclin D1, CD10, CD21, CD23, BCL2, BCL6, SOX11, Ki-67^b with or without
 - Cell surface marker analysis by flow cytometry with peripheral blood and/or biopsy specimen: kappa/lambda, CD19, CD20, CD5, CD23, CD10, CD200
- TP53 sequencing^c

^c TP53 mutation has been associated with poor prognosis in patients treated with conventional therapy, including transplant. Clinical trial is strongly recommended for these patients. TP53 by IHC is not a proven surrogate for TP53 mutation status or del(17p) status.

Rodrigues J et al. *Br J Haematol*. 2020;191(5):796-805.

MARGINAL ZONE LYMPHOMA



Nodal marginal zone lymphoma (NMZL)

- peripheral lymphadenopathy, often stage III or IV, often head & neck
- bone marrow involvement in ~1/3rd of cases
- does not involve extranodal sites or (rarely) peripheral blood
- marked splenomegaly uncommon
- may be associated with IgM monoclonal gammopathy (~10%)
- may be associated with autoimmune disease or hepatitis C infection

NODAL MARGINAL ZONE LYMPHOMA

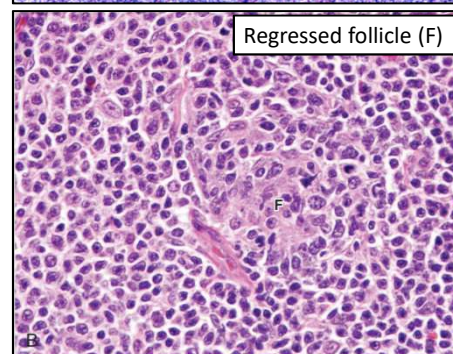
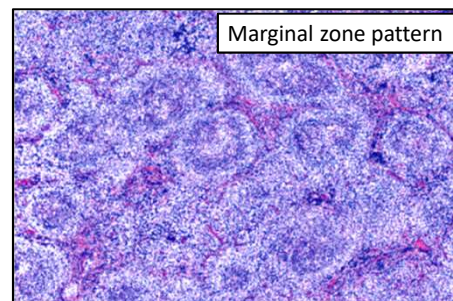
Challenges in diagnosis

Multiple architectural patterns

- marginal zone pattern (cells surrounding secondary follicles)
- follicular colonization pattern (neoplastic invasion of GCs)
- vaguely nodular pattern with absent/atrophic GCs
- diffuse pattern

Variable cytologic appearance

- centrocyte-like
- monocytoid
- plasmacytoid
- admixed larger, centroblast-like cells



NODAL MARGINAL ZONE LYMPHOMA

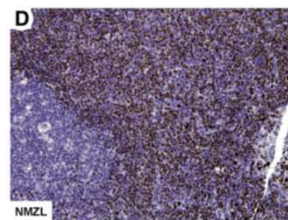
Challenges in diagnosis

Nodal marginal zone lymphoma	
CD20	+
CD5	- (~15% +)
CD10	-
BCL6	-
CD43	up to 50%
BCL2	+
CD23, CD21	negative in lesional cells but helpful to ID follicular dendritic cells
Plasmacytoid cells	
MUM-1	subset +
cytoplasmic light chain	restricted

Myeloid cell nuclear differentiation antigen (MNDA)

Table 1 MNDA expression analysis in extramedullary tissues involved by lymphoma

Diagnosis	Number MNDA+	Percent MNDA+
FL, grade 1-2	3/69 ^a	4.3%
FL, grade 3	3/41	7.3%
DLBCL	2/61	3.3%
NMZL	16/24	66.7%
EMZL	27/44	61.4%
SMZL	5/21	23.8%
Splenic diffuse red pulp small B-cell lymphoma	1/1	100%
CLL/SLL	4/31	12.9%
MCL	9/140	6.4%
LPL	2/8	25.0%



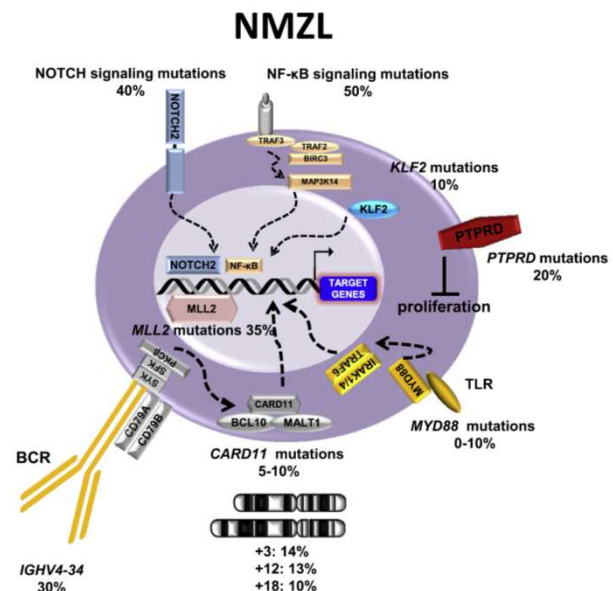
- myelomonocytic cells
- non-neoplastic B cells in spleen, in marginal zone distribution

Metcalf RA et al. *Human Pathology*. 2014;45, 1730-1736.
Angelopoulos MK et al. *Leukemia & Lymphoma*. 2014;55:6, 1240-1250.

NODAL MARGINAL ZONE LYMPHOMA

Challenges in diagnosis

- majority of recurrent genetic aberrancies are non-specific
 - trisomy 3
 - trisomy 12
 - trisomy 18
 - *NOTCH2* mutations
 - *NF-κB* signaling mutations
- *PTPRD* mutations appear to be specific for nodal MZL (~20% cases)

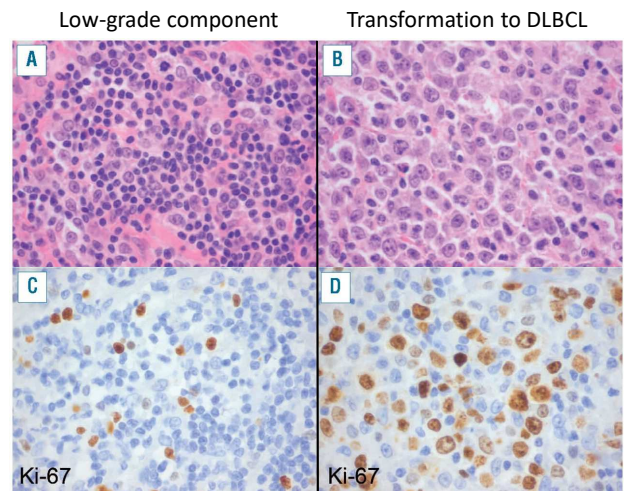


Spina V et al. *Best Practice & Research Clinical Haematology*. 2017;30:Se12.

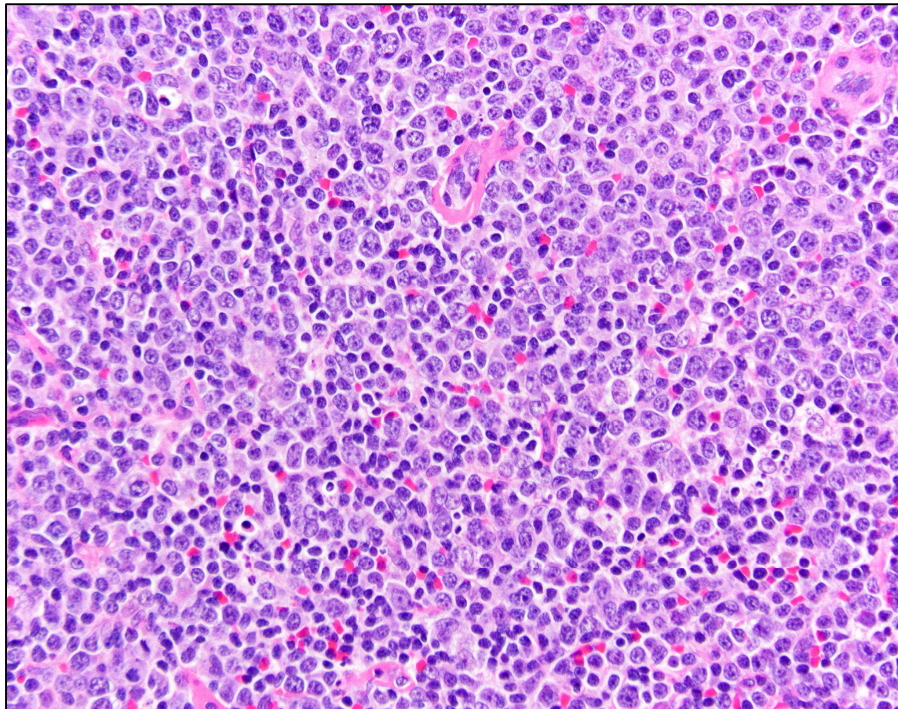
NODAL MARGINAL ZONE LYMPHOMA

Transformation

- transformation to DLBCL has been described in 15% of patients with NMZL at median 4.5 years from diagnosis
- caveat: morphologic distinction between NMZL and DLBCL can be challenging
 - NMZL can show an increased number of centroblasts, particularly in cases with follicular colonization
 - NMZL is not graded; higher proportion of large-sized cells does not definitively correlate with outcome
 - WHO recommends reserving diagnosis of DLBCL for cases with sheets of centroblasts



van den Brand M et al. *Hematologica*. 2013;98(7):1003-1013.

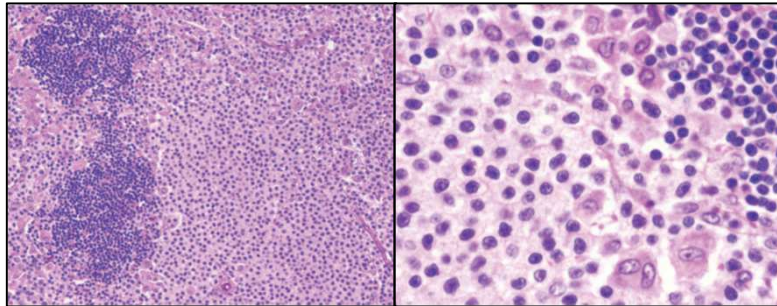


NODAL MARGINAL ZONE LYMPHOMA

Differential diagnosis

- 2° nodal involvement by EMZL or SMZL
 - exclusion requires careful clinical and radiologic correlation
 - large numbers of monocytoid cells and well-preserved reactive follicles favor nodal spread from an undetected EMZL

2° nodal involvement by EMZL



Jaffe ES. Chp 21. Hematopathology 2nd ed. 2016.

NODAL MARGINAL ZONE LYMPHOMA

Differential diagnosis

- lymphoplasmacytic lymphoma
 - similarities include IgM monoclonal gammopathy, absence of CD5/CD10 expression, plasmacytoid differentiation
 - *MYD88 L265P* mutations are found in >90% LPL cases but in only 0-10% of NMZL
- follicular lymphoma
 - NMZL with extensive follicular colonization can be difficult to distinguish from follicular lymphoma
 - in NMZL, the infiltrating NMZL cells will be BCL2+ whereas the residual follicular cells express BCL6, CD10 and stathmin but are negative for BCL2
- reactive conditions
 - marginal zone hyperplasia (reactive marginal zone cells are BCL2+ so this does not help distinguish)
 - hyperplasia of monocytoid B-cells (toxoplasmosis or other infections)

Often a diagnosis of exclusion given absence of defining immunophenotypic or genetic markers

Jaffe ES. Chp 21. Hematopathology 2nd ed. 2016.

HAIRY CELL LEUKEMIA

Nodal and extranodal involvement

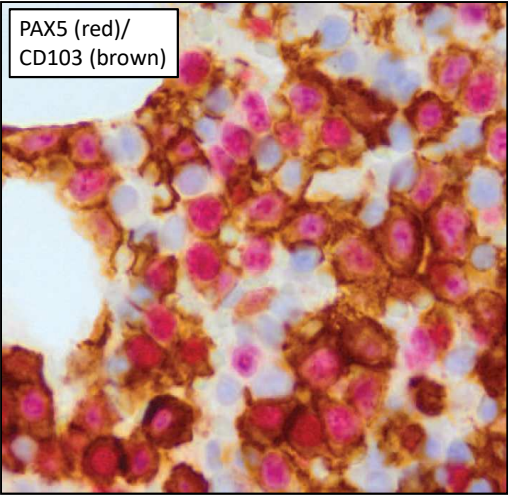
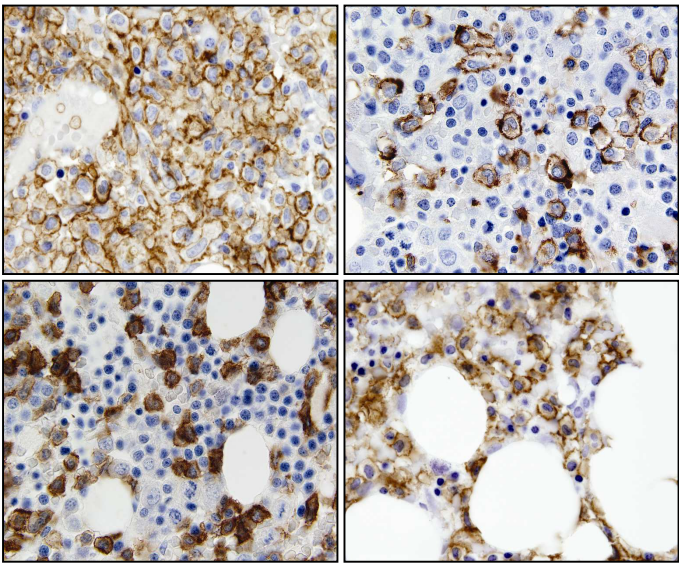
- lymph node involvement considered uncommon
 - often associated with advanced disease and splenectomy
- extranodal involvement is rare
 - can occur at presentation or in patients with longstanding disease
 - variable anatomic sites
 - lytic bone lesions
 - central nervous system
 - skin
 - breast
 - salivary gland
 - pleura

Cortazar JM et al. *Histopathology*. 2017;71, 112–124.

Hairy cell leukemia	
CD20	+
CD5	-
CD10	-
CD23	-
CD43	-/+
LEF1	-
CD200	+
FMC7	+
cyclin D1	+ (weak)
SOX11	-
light chain	+
TRAP	+
CD11c	+ (bright)
CD25	+
CD103	+
DBA.44	+
CD123	+
T-bet	+
Annexin A1	+
CD1d	+

HAIRY CELL LEUKEMIA

CD103

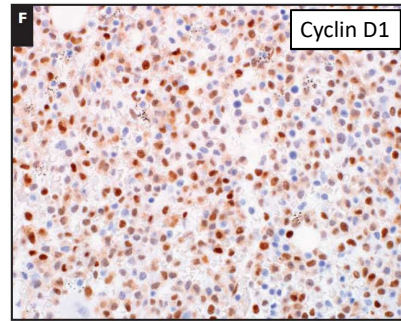


Rozenvald IB et al. *Arch Pathol Lab Med*. 2017;141(6):837-840.
Morgan EA et al. *Am J Clin Pathol*. 2013;139:220-230.

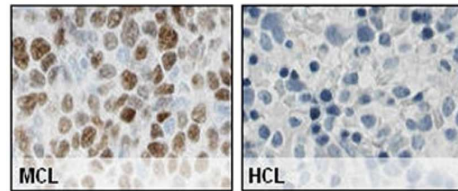
HAIRY CELL LEUKEMIA

IHC pitfalls

- cyclin D1+
 - weaker, less uniform than mantle cell lymphoma
 - not associated with t(11;14)
- SOX11-negative
 - earlier literature suggested HCL was SOX11+
 - this was using polyclonal SOX11 antibody
 - HCL does not express SOX11 when using a monoclonal antibody
- up to 25% express CD10
 - BCL6-negative
- rarely CD5+

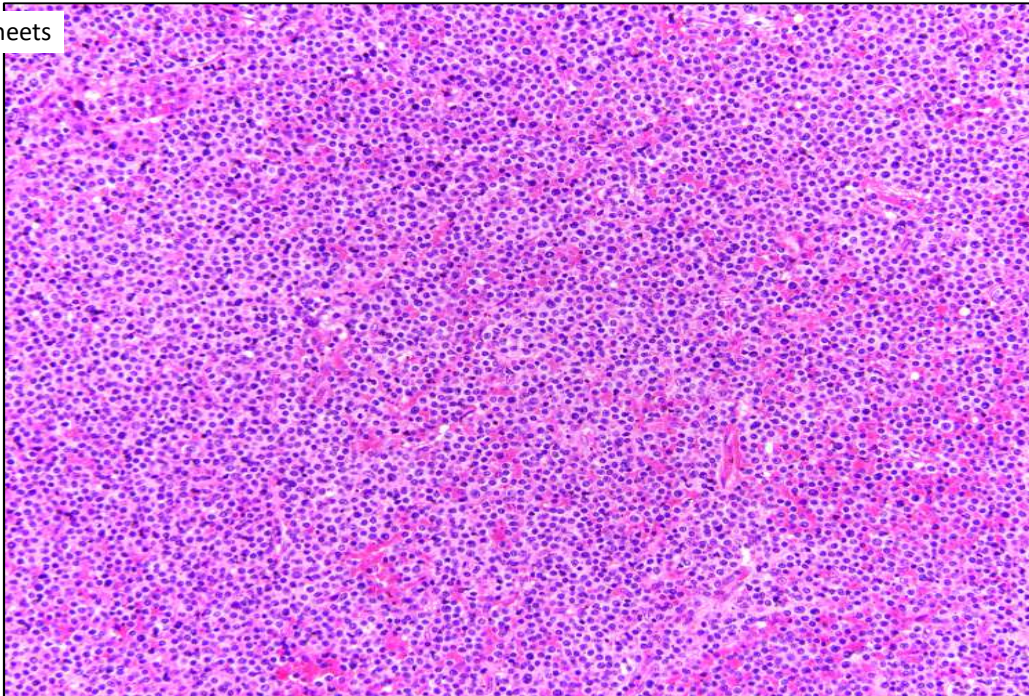


Monoclonal SOX11 (C-terminal peptide):

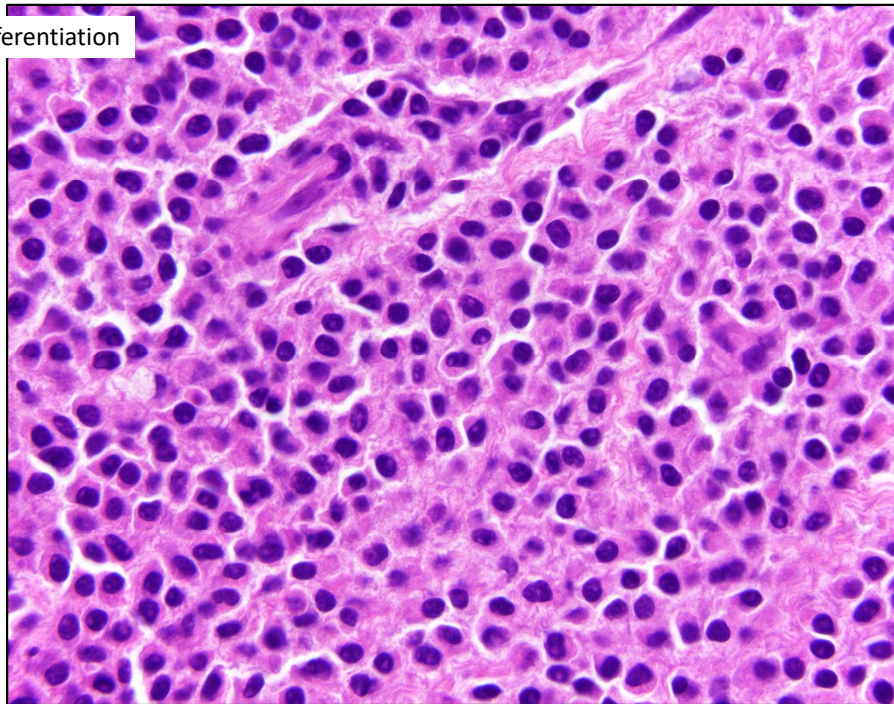


Jasionowski TM et al. *Am J Clin Pathol*. 2003;120:228-235.
Basso K et al. *J Exp Med*. 2004;199(1):59-68.
Cortazar JM et al. *Histopathology*. 2017;71, 112-124.
Bosch F et al. *Br Haematol*. 1995;91:1025-30.
Miranda RN et al. *Mod Pathol*. 2000;13(12):1308-1314.
Sherman MJ et al. *Am J Clin Pathol*. 2011;136:390-399.
Díctor M et al. *Haematologica*. 2009;94:1563-1568.
Nordström L et al. *BMC Cancer*. 2012;12:269.

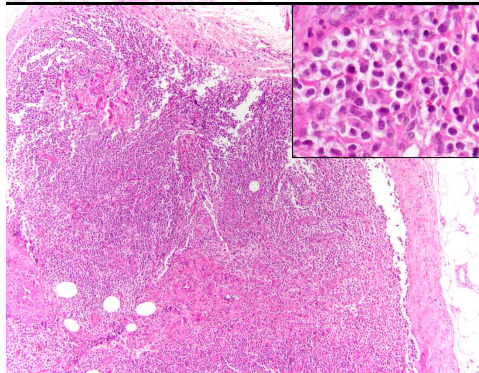
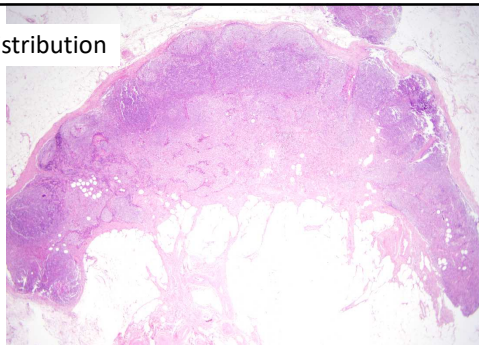
Diffuse sheets



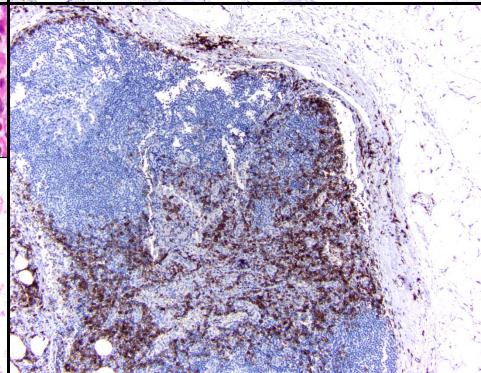
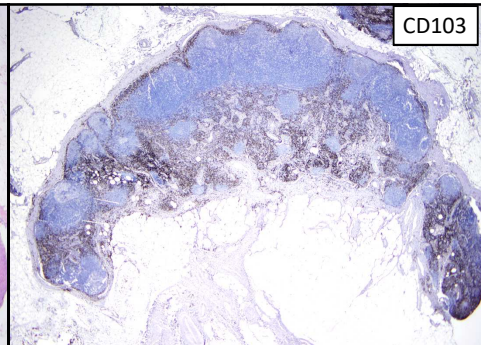
Plasmacytic differentiation



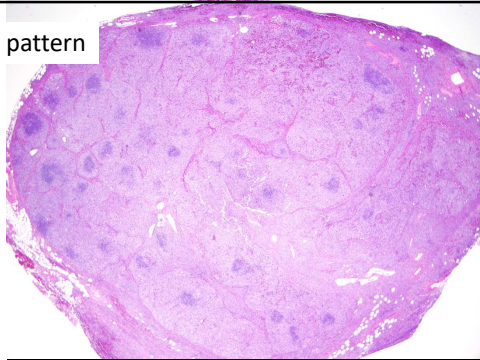
Sinusoidal distribution



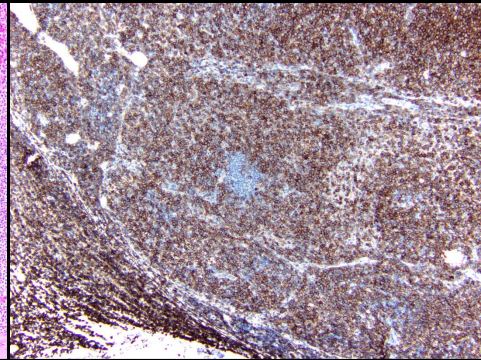
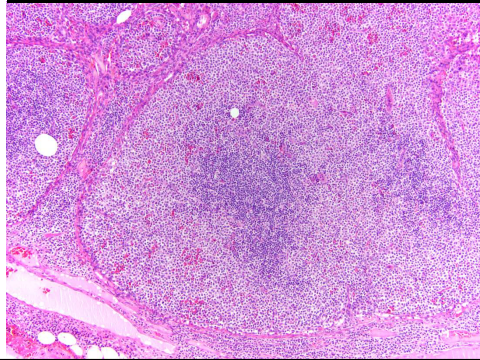
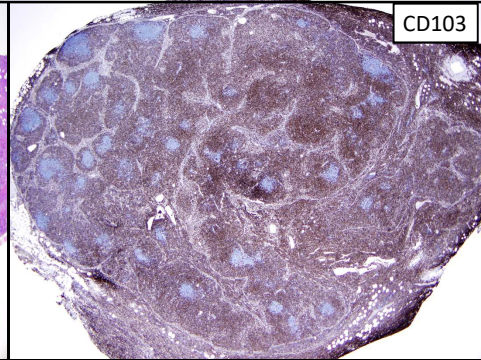
CD103



Pseudo-MZL pattern

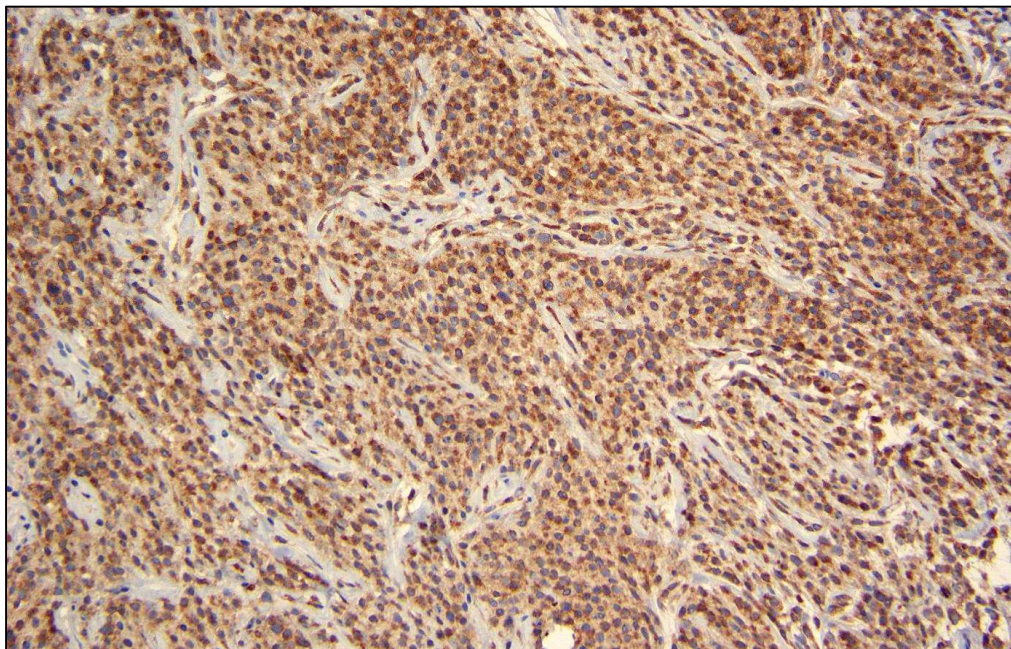


CD103



Cortazar JM et al.
Histopathology, 2017;
71, 112-124.

BRAF
V600E



SUMMARY

Transformation caveats

- expanded proliferation centers can be misinterpreted as Richter transformation
- HRS cells in CLL/SLL without mixed inflammatory background does not equal Richter transformation per WHO criteria, but should be reported
- necrosis in CLL/SLL does not always = transformation; consider HSV lymphadenitis
- blastoid or pleomorphic MCL, particularly *de novo*, can resemble large cell or immature process; perform cyclin D1 liberally in work-up of B-cell entities
- NMZL may have many centroblast-like cells, particularly in the setting of follicular colonization; diagnosis of DLBCL is reserved for sheets of centroblast-like or large-sized cells

SUMMARY

IHC pitfalls and helpful hits

- see tables in appendix for typical IHC profiles in small B-cell lymphomas, and for frequency of “aberrant” expression of CD5 and CD10 in B-cell neoplasms
- SOX11 is helpful for diagnosis of MCL but care is needed when choosing antibody clone
- cyclin D1 is not only positive in MCL but also plasma cell myeloma with t(11;14) which are often IgM+ and CD20+; HCL (weak); and proliferation centers of CLL/SLL
- p53 IHC is not a proven surrogate for *TP53* mutation or del(17p) status
- myeloid cell nuclear differentiation antigen (MNDA) is positive in approximately 60% of NMZL and EMZL but is also expressed in smaller subset of most other small B-cell lymphomas



IMMUNOHISTOCHEMISTRY (APPENDIX)

Table 14-7 Classic Immunophenotypic Profile of B-Chronic Lymphoproliferative Neoplasms

Disorder	Slg	CD20	CD22	CD23	CD25	CD5	FMC7	CD11c	CD10	CD79b	CD103	CD200	LEF1	Cyclin D1	SOX11
CLL/SLL	w	w	w	+	–	+	–/w	w	–	–/w	–	+ bright	+	–	–
PLL	+	+	+	–	–	v	v	–	–	+	–	–/+	ND	–	ND
MCL	+	+	+	–, w	–	+	+	–	–	+	–	–	–	+	+
FL	+	+	+	–	–	–	+	–	+	+	–	+ dim/mod	–	–	–
SMZL	+	+	+	–	–	v	+	v	v	+	–	+ dim/mod	ND	–	–
HCL	+	+	+	–	+	–	+	+	s	–	+	+ bright	ND	+s	–
LPL	+/CIg	+	+	–	–	–	–	–	–	+	–	+/- dim	–	–	–

References 1, 36, 40, 68, 71, and 199-203.

CIg, cytoplasmic immunoglobulin; CLL/SLL, chronic lymphocytic leukemia/small lymphocytic leukemia; FL, follicular lymphoma; HCL, hairy cell leukemia; LPL, lymphoplasmacytic; MCL, mantle cell lymphoma; ND, not done; PLL, polymorphous leukemia; +s, subset of cases positive; SMZL, splenic marginal zone lymphoma, lymphoma; v, variable expression; w, weakly expressed

Immunophenotype	Type of Mature B-Cell Lymphoma	Approximate % Staining	Source, y
CD5 ⁺ /CD10 ⁻	MCL	93–95	Dorfman and Shahsafaei, ⁶ 1997; Gualco et al, ⁷ 2010
	SLL/CLL	80–92	Kurec et al, ⁴ 1992; Geiler et al, ⁵ 1991
	LPL	9–43	Hunter et al, ²⁰ 2005; Morice et al, ²¹ 2009
	MZBCL		
	Splenic MZBCL	20	Gimeno et al, ²⁴ 2005; Matutes et al, ²⁵ 2008
	Nodal MZBCL	8.6	Jaso et al, ²² 2013
	MALT lymphoma	1	Jaso et al, ²³ 2012
	De novo DLBCL NOS	10	Tagawa et al, ²⁷ 2005
	BL	100	Dogan et al, ³⁷ 2000
	FL (low grade)	80	Eshoa et al, ³¹ 2001
CD10 ⁺ /CD5 ⁻	DLBCL, NOS	10–40	Colomo et al, ⁴⁶ 2003; Berglund et al, ⁴⁷ 2005; Visco et al, ⁴⁸ 2012
	HCL	10–20	Jasionowski et al, ⁴³ 2003; Gupta et al, ⁴⁴ 2015
	FL (grade 3)	17	Eshoa et al, ³¹ 2001
	MCL	0–7	Akhter et al, ⁴⁵ 2015
	MZBCL		
CD5 ⁻ /CD10 ⁻	MALT lymphoma	>99	Jaso et al, ²³ 2012
	Nodal MZBCL	92	Jasoco et al, ²² 2013
	Splenic MZBCL	80	Gimeno et al, ²⁴ 2005; Matutes et al, ²⁵ 2008
	LPL	57–91	Hunter et al, ²⁰ 2005; Morice et al, ²¹ 2009
	HCL	80–90	Jasionowski et al, ⁴³ 2003; Gupta et al, ⁴⁴ 2015
	DLBCL, NOS	50–70	Tagawa et al, ²⁷ 2005; Colomo et al, ⁴⁶ 2003; Berglund et al, ⁴⁷ 2005; Visco et al, ⁴⁸ 2012
	FL (grade 3)	83	Eshoa et al, ³¹ 2001
	FL (low grade)	20	Eshoa et al, ³¹ 2001
	SLL/CLL	8–20	Kurec et al, ⁴ 1992; Geiler et al, ⁵ 1991

Abbreviations: BL, Burkitt lymphoma; CLL, chronic lymphocytic leukemia; DLBCL, diffuse large B-cell lymphoma; FL, follicular lymphoma; HCL, hairy cell leukemia; LPL, lymphoplasmacytic lymphoma; MALT, mucosa-associated lymphoid tissue; MCL, mantle cell lymphoma; MZBCL, marginal zone B-cell lymphoma; NOS, not otherwise specified; SLL, small lymphocytic lymphoma.

Wang HY et al. Arch Pathol Lab Med. 2017;141:1236–1246.