

## ***Hot topics in myeloid neoplasms***

*Valentina Nardi, MD*

*Associate Professor of Pathology, Harvard Medical School  
Assistant pathologist, Massachusetts General Hospital*

### ***Outline***

- ❖ *Premalignant clonal myeloid proliferations*
- ❖ *Myeloid neoplasms with germline predisposition*
- ❖ *Mutations and MDS diagnosis*
- ❖ *What is new in the genetic testing workflow for MPN, eosinophilia, AML*
- ❖ *MRD for AML*

## Premalignant clonal myeloid proliferations

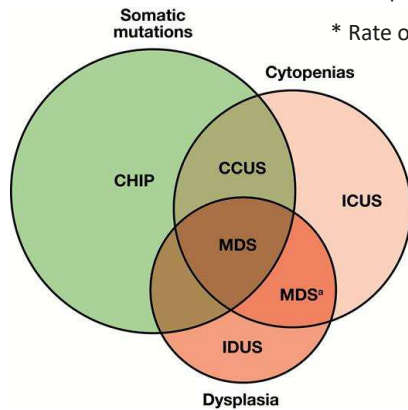
**ARCH / CH/ CHIP** = somatic mutations of genes frequently mutated in hematologic malignancies (*DNMT3A* > *ASXL1* > *TET2* > *JAK2* > *PPM1D* > *SF3B1* > *SRSF2* > *TP53*..) with a VAF  $\geq 2\%$ , with no clinical/morphologic evidence of a hematologic malignancy

\* Accompanies aging (10% over age 70, 20% over age 90)

\* Rate of progression to hematologic neoplasm  $\sim 0.5\%-1\%$  per year

\* Rate of progression is proportional to the size (VAF) of the somatic clone (the higher the VAF, the higher the risk)

\* *TP53* and *PPM1D* mutations are associated with increased risk of therapy-related myeloid neoplasm, and lymphoma post ASCT



Clonal hematopoiesis of indeterminate potential [CHIP]  
 Idiopathic cytopenia of undetermined significance [ICUS]  
 Clonal cytopenias of undetermined significance [CCUS]  
 Idiopathic dysplasia of undetermined significance [IDUS]  
 Myelodysplastic syndrome [MDS]  
 Clonal hematopoiesis [CH]  
 Age-related clonal hematopoiesis [ARCH]

Nardi V et al, AJCP 2019

Jaiswal S., et al., N Engl J Med 2014; Genovese G. et al, NEJM 2014;

	Cytopenia	Dysplasia	CH	VAF	No. of Variants	Genes	Risk
Normal	-	-	-	-	-	-	-
CHIP	-	-	+	$\geq 2\%$ but $< 20\%$	Rarely $> 2$	<i>TET2</i> , <i>DNMT3A</i> , <i>ASXL1</i> , <i>SF3B1</i> , <i>PPM1D</i>	1%/y
ICUS	+	-	-	-	-	-	low
CCUS	+	-	+	$> 10\%$	Often $> 1$	<i>SRSF2</i> , <i>IDH1</i> , <i>NRAS</i> , <i>RUNX1</i> , <i>U2AF1</i> , <i>BCOR</i> , <i>EZH2</i> , <i>STAG2</i>	10%/y
MDS	+	+	+/-	$> 10\%$	On average $\geq 2$	-	-
IDUS	-	+	-	-	-	-	-

## Premalignant clonal myeloid proliferations: CH vs AML MRD

Genetic abnormality	Type	Usually cleared after successful therapy	Persistence after therapy associated with adverse outcome
<i>RUNX1-RUNX1T1</i> , <i>CBFB-MYH11</i> , <i>PML-RARA</i>	AML-related	Yes	Yes
<i>NPM1</i>	AML-related	Yes	Yes
<i>KMT2A</i> rearrangement, <i>DEK-NUP214</i> , <i>BCR-ABL1</i>	AML-related	Unknown	Unknown
<i>NRAS/KRAS</i>	AML-related	Yes	Yes
<i>FLT3-ITD/FLT3-TKD</i>	AML-related	Yes (but may be lost at relapse or acquired at relapse of previously <i>FLT3</i> wild-type AML)	Unknown
<i>KIT</i>	AML-related	Yes	Yes
<i>PTPN11</i>	AML-related	Yes	Yes
<i>IDH1/IDH2</i>	CH (potentially AML-related)	Variable	Yes
<i>DNMT3A</i>	CH	Usually not	No
<i>ASXL1</i>	CH	Variable	No
<i>TET2</i>	CH	Usually not	No

CH may persist at AML  
 Remission: ddx with MRD!

Adapted from Hasserjian RP, et al., Blood 2020

## Myeloid neoplasms with germline predisposition

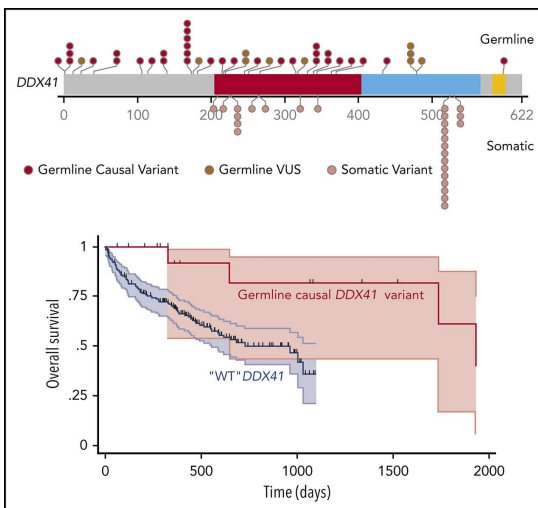
- Increased recognition; relevance for bone marrow donor selection
- Many of the genes mutated in the germline can also be mutated as acquired events in MDS/AML; importance of family and personal history and awareness
- Skin fibroblasts, nails, hair for germline testing

Mutated gene	Region	Inheritance	1st report	Median age at diagnosis (range), years	Low platelets	Other organ dysfunction	Type of neoplasm	Risk of HM
<i>CEBPA</i>	19q13.1	AD	2004	25 (2–46)	no	no	AML	100%
<i>DDX41</i>	5q35.3	AD	2015	62 (40–85)	no	no	AML, MDS, rarely CML, CMML, lymphoma, myeloma	?%
<i>RUNX1</i>	21q22.12	AD	1999	39 (7–53)	yes	no	AML, MDS, rarely CMML, T-ALL, hairy-cell leukemia	40%
<i>ANKRD26</i>	10p12.1	AD	2011	38 (1–84)	yes	no	AML, MDS, rarely CML, CMML, CLL	8%
<i>ETV6</i>	12p13.2	AD	2015	uncertain	yes	no	B-ALL, AML, MDS, CMML, myeloma, PV, solid tumors	8%
<i>GATA2</i>	3q21.3	AD	2010	20 (<1 to 78)	no	yes	AML, MDS, CMML, aCML	80%
<i>SAMD9/SAMD9L</i>	7q21.2	AD	2016	uncertain	yes	yes	MDS, AML	?%

-Bone marrow failure syndrome  
-Telomeropathies  
-JMML

Adapted from Geyer J. T. , Myeloid Neoplasms with Germline Predisposition. Pathobiology. 2019;

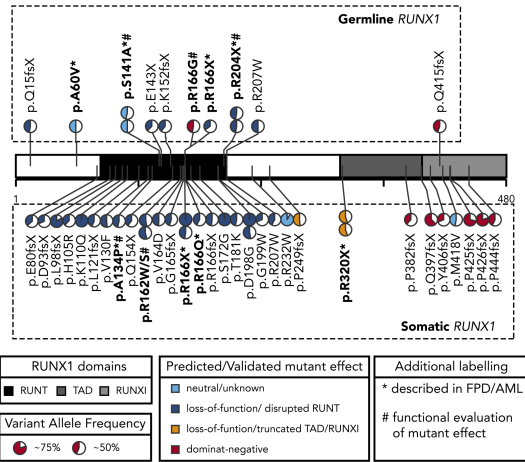
## Myeloid neoplasms with germline *DDX41* mutations



Sebert M., et al, Blood 2019

- DEAD-box helicase 41 (*DDX41*), essential for cell growth and viability of hematopoietic stem and progenitor cells
- ~1-4% of myeloid neoplasms
- Antecedent cytopenias, particularly leukopenia
- Male gender
- Average age of MDS/AML onset in mutation carriers is notably older at 65 years
- Most germline mutations are truncating (or M1 or codon R525)
- Most common somatic mutation is a second *DDX41* mutation, usually missense
- HSCT from *DDX41* mutation carriers may promote donor cell leukemias
- Lenalidomide has been suggested as an effective treatment strategy for myeloid malignancies with *DDX41* mutations [and without del(5q)]

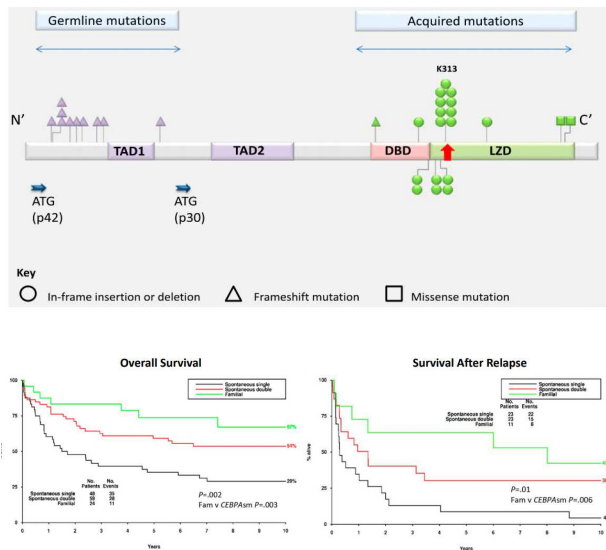
## Myeloid neoplasms with germline *RUNX1* mutations: familial platelet disorder (FPD) with associated myeloid malignancy (FPDMM, also referred to as FPD/AML)



- RUNX1* encodes a TF that is a master regulator of hematopoiesis
- Germline mutations in *RUNX1* occur in ~15--30% of patients with AML and *RUNX1* mutations
- Variable clinical presentation, with mostly mild to moderate bleeding tendency since childhood
- Platelet count are often normal but there platelet dysfunction
- Germline *RUNX1* mutations encompass partial and whole gene deletions and frameshift, stop-gain, and missense mutations
- Median age of onset of MDS/AML is 33 ys
- Somatic mutations in *RUNX1* are frequently observed in leukemic progression; *NRAS* mutations are also common

Simon et al, *Blood*. 2020

## Myeloid neoplasms with germline *CEBPA* mutations



- CEBPA* is a transcription factor involved in the control of myeloid progenitor differentiation and proliferation
- Germline mutations occur typically in the N-terminal
- AML develops with acquisition of a somatic *CEBPA* mutation in the C-terminal region
- Patients usually develop AML as children and young adults (a median age of 25 years)
- Approximately 10% of AML cases with biallelic mutations represent a germline and a somatic mutation
- Somatic *CEBPA* mutations appear unstable throughout the disease course, with novel independent clones frequently identified at recurrence
- Favorable prognosis

Tawana K, et al, *Blood* 2015

## Mutations and diagnosis of MDS

### 1. MDS-defining cytogenetic aberrations in the absence of significant morphologic dysplasia

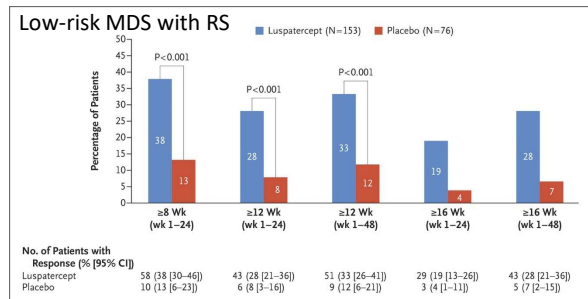
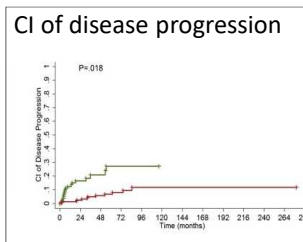
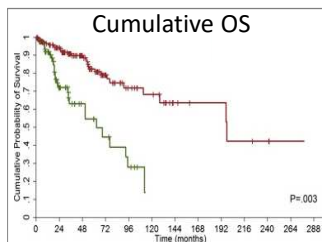
Chromosomal abnormality	Frequency	
	MDS overall	Therapy-related MDS
<b>Balanced</b>		
t(11;16)(q23.3;p13.3)		3%
t(3;21)(q26.2;q22.1)		2%
t(1;3)(p36.3;q21.2)	1%	
t(2;11)(p21;q23.3)	1%	
inv(3)(q21.3;q26.2)/t(3;3)(q21.3;q26.2)	1%	
t(6;9)(p23;q34.1)	1%	

Chromosomal abnormality	Frequency	
	MDS overall	Therapy-related MDS
<b>Unbalanced</b>		
<del>Gain of chromosome 8<sup>a</sup></del>	10%	
Loss of chromosome 7 or del(7q)	10%	50%
del(5q)	10%	40%
<del>del(20q)<sup>a</sup></del>	5–8%	
<del>Loss of Y chromosome<sup>a</sup></del>	5%	
Isochromosome 17q or t(17p)	3–5%	25–30%
Loss of chromosome 13 or del(13q)	3%	
del(11q)	3%	
del(12p) or t(12p)	3%	
del(9q)	1–2%	
idic(X)(q13)	1–2%	

Adapted from Swerdlow SH, et al.: WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues (Revised 4th edition).

## Mutations and diagnosis of MDS

### 2. SF3B1 mutations and 5-15% ring sideroblasts

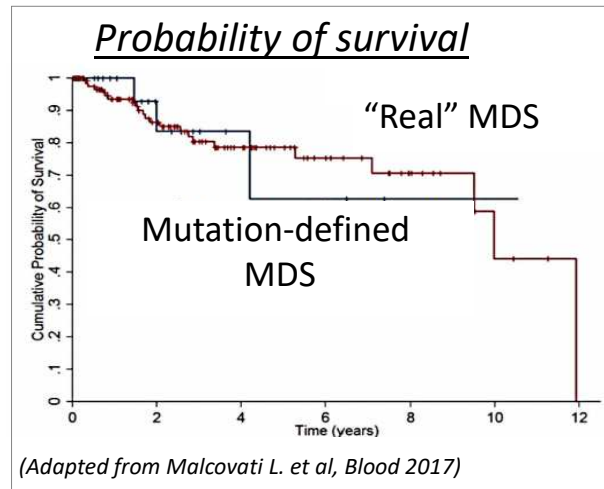


Adapted from Swerdlow SH, et al.: WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues (Revised 4th edition) and Malcovati L. et al, Blood 2015; Fenaux P. et al., NEJM 2020

## Mutations and diagnosis of MDS

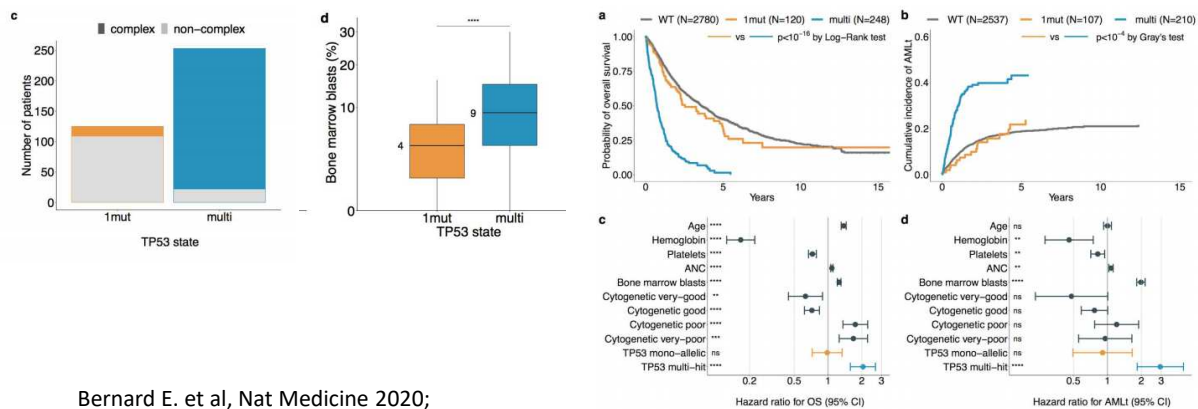
3. Presence of specific **mutation patterns**, “**high**” VAF (>10% of ) and unexplained **cytopenias**  $\sim$ /= MDS

- **Spliceosome mutations** (SF3B1, SRSF2, U2AF1, ZRSR2)
- **DNMT3A/TET2/ASXL1** (“DTA” mutation) + another mutation



## TP53 Mutations in MDS

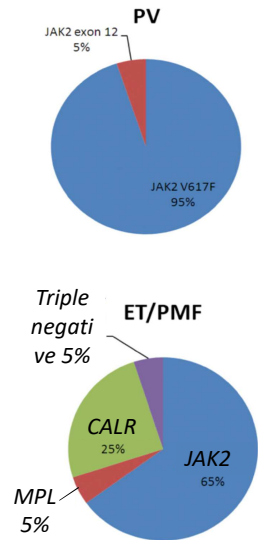
- **Biallelic** not mono-allelic **TP53 mutations** are associated with complex karyotype, high risk presentation, poor outcome, resistance to conventional therapy.
- Biallelic/two hits: mutation, deletion, or loss of heterozygosity



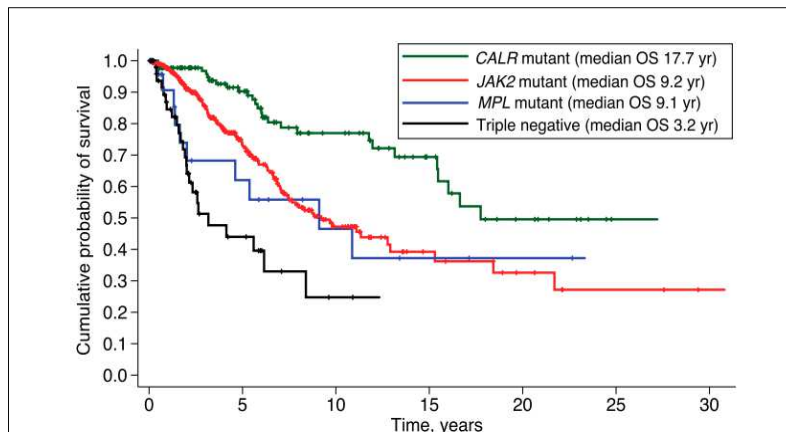
Bernard E. et al, Nat Medicine 2020;

## **Genetic testing for diagnosis and classification of MPN (except eosinophilias)**

- ❑ **Karyotype** (clonality, progression..)
- ❑ **BCR-ABL1; ABL1 mutations**
- ❑ **JAK2 V617F** (and exon 12 mutations), **MPL** and **CALR**. Broader myeloid mutation panel if triple-negative.
- ❑ **CSF3R** in suspected chronic neutrophilic leukemia



## **Risk stratification of MPN according to driver mutations**



**N=617**  
**PMF**

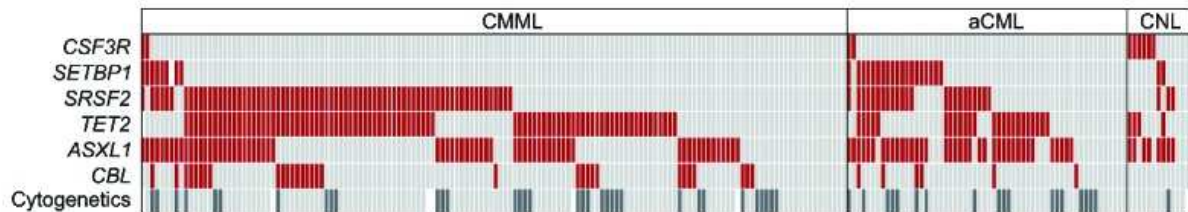
*Elisa Rumi et al, Blood 2016*

**CALR mutations:** favorable prognosis, indolent clinical course

**JAK2/MPL mutations:** intermediate prognosis. **Increased risk of thrombosis**

**Triple neg:** unfavorable prognosis; high risk of transformation to AML

## Overlap in mutational landscape of CMML, aCML and CNL



**CSF3R mutations** in 90% of patients with **CNL** (truncation mutations → constitutive overexpression of the receptor and ligand hypersensitivity; membrane proximal mutation → constitutive activation of the receptor)

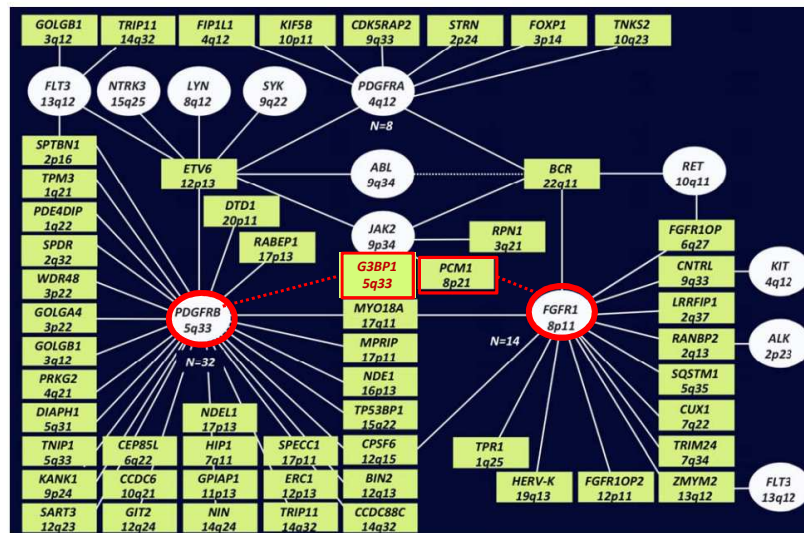
*Meggendorfer et al, Haematologica 2014*

## Genetic testing for diagnosis and classification of eosinophilias

- ☐ **Karyotype** (clonality → Chronic eosinophilic leukemia NOS)
- ☐ **BCR-ABL1** (FISH, RT-PCR, karyotype)
- ☐ **FIP1L1-PDGFR** by FISH (cryptic)  
\*Consider PDGFRB FISH, FGFR1 FISH, Fusion panel
- ☐ **Broad myeloid-mutation panel** (prove clonality) including **KIT** (exclude mastocytosis)



## Fusion genes associated with myeloid/lymphoid neoplasms with eosinophilia



Reiter A, Gotlib J, Blood 2018

Jan M. et al, A cryptic imatinib-sensitive G3BP1-PDGFRB rearrangement in a myeloid neoplasm with eosinophilia. Blood Advances 2020

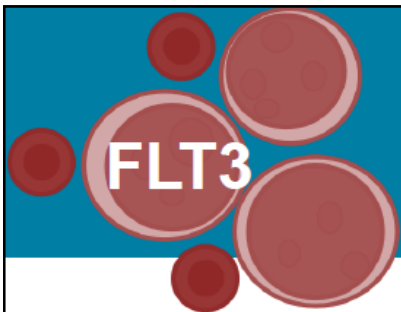
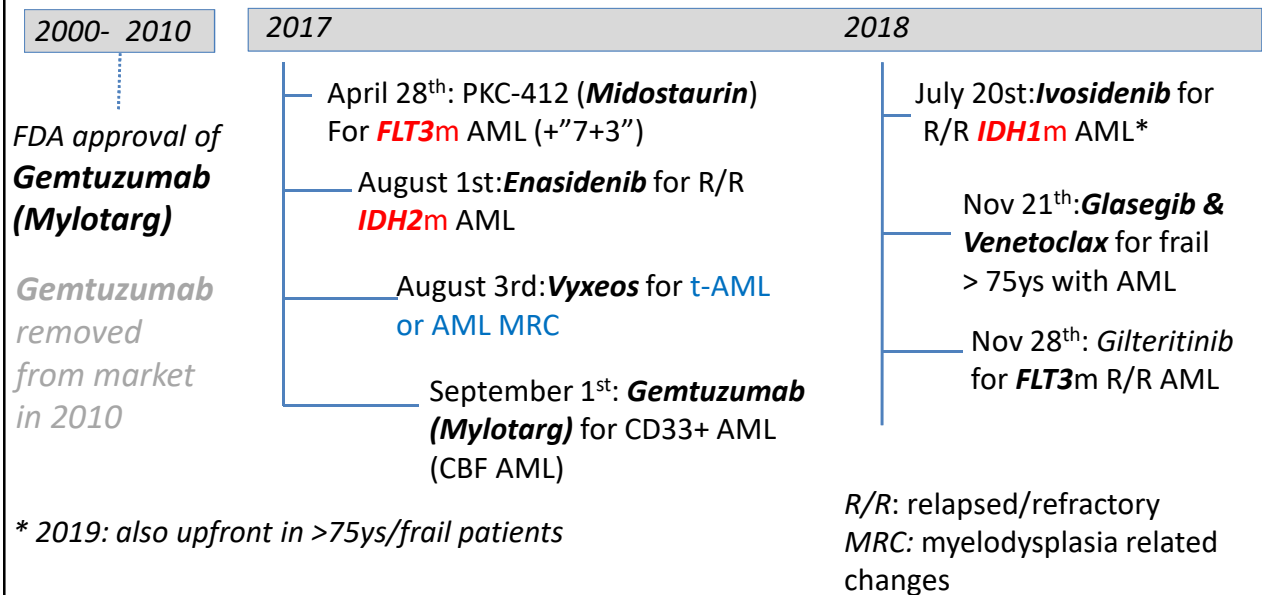
Kasbekar M et al. Targeted FGFR inhibition results in a durable remission in an FGFR1-driven myeloid neoplasm with eosinophilia. Blood Adv 2020

## Genetic testing for diagnosis and classification of AML (CAP-ASH guidelines 2017)

- ☐ Karyotype
- ☐ FLT3 ITD and TKD\*, NPM1, CEBPA, RUNX1, IDH1/2\*
- ☐ May also perform **additional mutational analysis** (WT1, TP53, ASXL1..)
- ☐ KIT in core binding factor AML
- ☐ NUP98 rearranged AML (pediatric AML>>)

\* Need rapid Turnaround time

## Role of the molecular laboratory in the era of FDA approved drugs



## Role of molecular laboratory

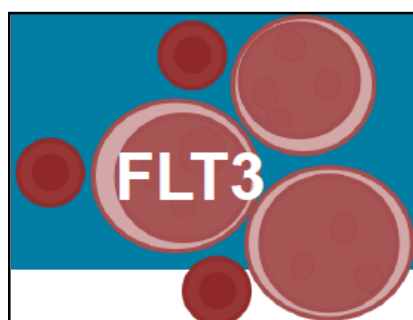
Test at AML diagnosis (and relapse):

### -FLT3 ITD

- fragment analysis
- NGS

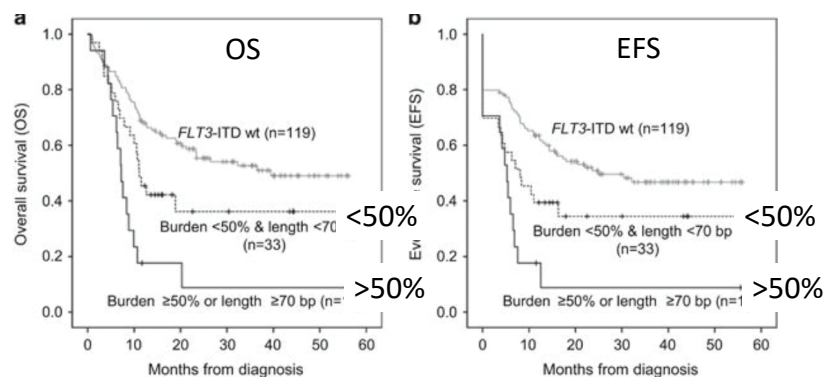
### -FLT3 TKD

- allele specific PCR
- Sanger sequencing
- NGS

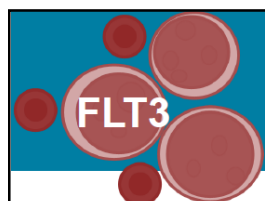


Part of ELN risk  
Stratification:  
ITDlow: AR<0.5  
ITDhigh: AR>0.5

## FLT3 ITD burden matters (allelic ratio (AR))



Kim Y, et al. Blood Cancer Journal 2015

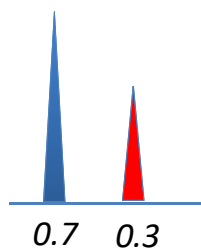


## FLT3 ITD AR controversies

### 1. How is it calculated?

VAF: ratio AF ITD/ AF (ITD+WT)

■ FLT3 wt  
■ FLT3 ITD

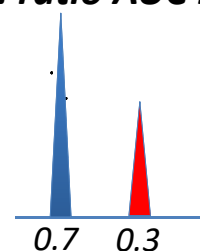


$$0.3/(0.7 + 0.3) = 0.3 \text{ VAF}$$

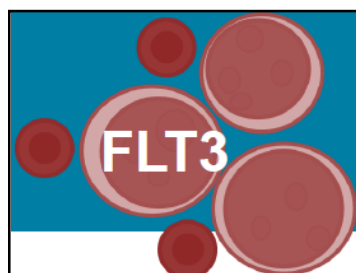
(NGS)

AR: ratio AUC ITD/ AUC WT

(sizing assay)



$$0.3/0.7 = 0.42 \text{ AR}$$



## ***FLT3 ITD: allelic ratio controversies***

### **2. What AR cutoff?**

Reference	Allelic ratio			Patients, N	Population
	Low	Intermediate	High		
Thiede C, et al.	$\leq 0.78$	NA	$> 0.78$	979	Adult, $> 18$ y
Gale RE, et al.*	$< 25\%$	25%-50%	$> 50\%$	1425	Adult, 18-60 y
Meshinchi S, et al.	$\leq 0.4$	NA	$> 0.4$	630	Pediatric, 0-21 y
Linch DC, et al.*	$< 25\%$	25%-50%	$> 50\%$	1609	Adult
→ Schlenk RF, et al.	$< 0.51$	NA	$\geq 0.51$	323	Adult, 16-62 y

<https://www.medscape.org/viewarticle/898477> transcript



## ***Role of molecular laboratory***

**Test at AML diagnosis for elderly and frail patients:**

-IDH1 codon 132 mutations

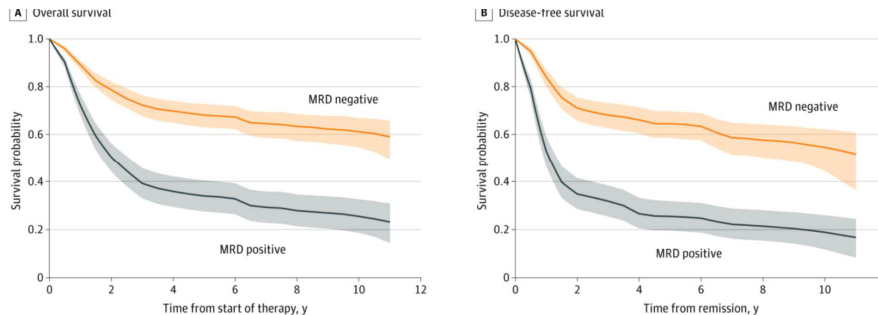
**Test at AML relapse or in refractory disease:**

-IDH1 codon 132

-IDH2 codon 140 and 172

## Minimal (Measurable) residual disease (MRD) in AML

- ❖ MRD should be standard of care
- ❖ MRD positivity after CR in a patient with AML is associated with a higher risk of relapse and shorter survival
- ❖ MRD in AML can be assessed using **MFC** and **PCR** approaches (*more in Dr. Annette Kim's presentation*)  
PCR ↔ RUNX1-RUNX1T1, CBFBMYH11, PML-RARA and mutations in NPM1.  
(While **NGS** may be applicable to another 40%-50% of AML patients, its use is still being standardized)
- ❖ The strongest evidence for MRD is in **core-binding AML**, but persistence of NPM1 mutation in remission by ultrasensitive techniques is also being used to make treatment decisions, though has not yet made its way into the guidelines.



Short N., et al. Association of Measurable Residual Disease With Survival Outcomes in Patients With Acute Myeloid Leukemia. *JAMA Oncology*, 2020; Schuurhuis GJ, et al. Minimal/measurable residual disease in AML: a consensus document from the European LeukemiaNet MRD Working Party. *Blood*. 2018;

## Conclusions

- ❖ Be aware of germline pathogenic mutations conferring increased risk of hematological malignancies
- ❖ Mutations in myeloid elements can represent clonal hematopoiesis, CCUS, MDS, AML MRD
- ❖ Need of rapid test results for patients with AML (FLT3, IDH1/2 at a minimum)
- ❖ Testing for cryptic gene fusions in patients with unexplained eosinophilia
- ❖ Minimal residual disease detection in AML is being adopted and used for treatment decisions

***Thank you!***